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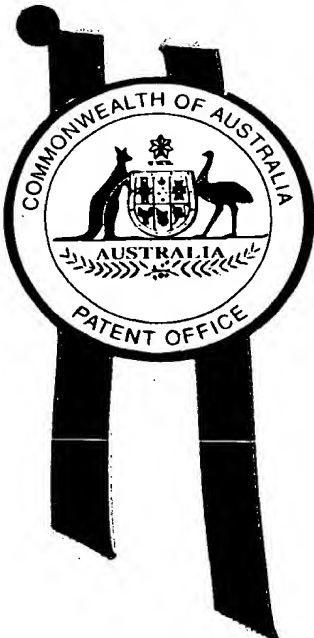
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I, KIM MARSHALL, MANAGER EXAMINATION SUPPORT AND SALES, hereby certify that the annexed is a true copy of the Provisional specification in connection with Application No. PO 9001 for a patent by MONASH UNIVERSITY filed on 8 September 1997.

I further certify that the above application is now proceeding in the name of METABOLIC PHARMACEUTICALS LTD pursuant to the provisions of Section, 113 of the Patents Act 1990.

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LTD

A U S T R A L I A
Patents Act 1990

PROVISIONAL SPECIFICATION
for the invention entitled:

Treatment of Obesity

The invention is described in the following statement:



- 1A -

TREATMENT OF OBESITY

FIELD OF THE INVENTION

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This invention relates to the treatment of obesity in animals. In particular, the invention relates to the treatment of obesity in humans, although it is to be understood that the present invention also extends to the treatment of obesity in non-human mammals, for example, for the improvement of meat qualities in farm animals used in food production.

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BACKGROUND OF THE INVENTION.

The critical role of human growth hormone (hGH) in postnatal growth in humans is well recognised. Less obvious is the impact of this hormone on the regulation of lipid and carbohydrate metabolism, due to lack of detailed molecular studies.

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It is well documented that the predominant form of hGH is a globular protein with a molecular weight of 22,000 daltons (22-KD) and consists of 191 amino acid residues in a single-chain, folded by 2 disulphide bonds with a small loop at the carboxyl terminus between residues 182 and 189. Recent crystallographic studies also show that the hGH molecule contains four anti-parallel α -helices which are arranged in a left-twisted, tightly-packed helical bundle¹. The concept that there are discrete functional domains within the hGH molecule responsible for specific metabolic actions of the hormone is generally accepted. The amino-terminus has been identified as the functional domain responsible for the insulin-like actions of the hGH molecule^{2,3}.

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Recombinant DNA technology opens the way to the large-scale commercial production of human growth hormone, and the recombinant hGH appears to have equivalent biological efficacies and pharmacokinetic properties^{4,5}. Current supply of this multiple-functional hormone no longer restricts the types and numbers of experimental therapies in humans and animals. The use of hGH for treatment of short stature in children and adults is well-established⁶. Therapeutic effects of hGH in female infertility have also been reported^{7,8}. Treatment of human obesity with hGH encounters a variety of problems. Evidence suggests that this multiple-functional hormone often simultaneously exerts *in vivo*, by various bioactive domains within the molecules, some adverse effects^{9,10}.

Regulation of lipid metabolism by GH was first described in 1959 by Raben & Hollenberg¹¹. The regulatory role of the hormone in lipid metabolism was subsequently supported by the body composition studies of GH-deficient and GH-treated humans^{12,13} and pigs^{14,15}. The findings of Gertner suggest that hGH is linked to adipose tissue distribution through a series of interactions known as the "GH-fat cycle"¹⁶. However, the molecular events transpiring to these biochemical and physiological changes remained largely unknown. The metabolic effects of GH on adipose and other tissues *in vivo* are variable and complex, apparently consisting of at least two components, an early insulin-like effect followed by a later more profound anti-insulin effect¹⁷. The results of the latter effect may include both a stimulation of lipolysis and an inhibition of lipogenesis. The anti-lipogenic effect of hGH has been substantiated with the demonstrations of the decrease of the expression of glucose transporter GLUT 4 in adipocytes¹⁸, the inhibition of the activity of acetyl-CoA carboxylase in adipose tissues^{19,20} and the reduction of glucose incorporation into lipid in both isolated cells and tissues^{21,22}.

In view of the multiple-functional effects of intact hGH and the problems encountered in clinical applications of the intact hormone, work leading to the present

invention has been directed to investigating whether hGH derivatives could be synthesised that retain the desired bioactivities and lack the unwanted side effects.

The structure-function studies of hGH with synthetic hormonal fragments have revealed that the carboxyl terminus of the hGH molecule appears to be the functional domain of the hormone for the regulation of lipid metabolism^{20,23} and it has now been shown that a synthetic peptide having a sequence based in the carboxyl terminal region reduces body weight gain and adipose tissue mass in a laboratory obese animal model.

SUMMARY OF THE INVENTION

The present invention concerns a method for treating obesity comprising administering an effective amount of a peptide which comprises the carboxyl-terminal sequence of a growth hormone. The treatment may be administered to any animal, including humans. The peptide may comprise the carboxyl-terminal sequence of human growth hormone or the growth hormone of a non-human mammalian specie. In one embodiment of the invention, the peptide comprises the carboxyl-terminal sequence of human growth hormone containing amino acid residues 177-191. In other embodiments of the invention, the peptide comprises a homologue, analogue, mutant, variant or derivative obtained by insertion, deletion or substitution of amino acids in, or chemical modification of, the native carboxyl-terminal sequence of human growth hormone or the growth hormone of a non-human mammalian species.

The present invention also concerns a pharmaceutical composition for use in the treatment of obesity comprising an effective amount of a peptide which comprises the carboxyl-terminal sequence of a growth hormone and one or more pharmaceutically acceptable carriers and/or diluents. The peptide may comprise the carboxyl-terminal sequence of human growth hormone or the growth hormone of a non-human mammalian specie. In one embodiment of the invention, the peptide

comprises the carboxyl-terminal sequence of human growth hormone containing amino acid residues 177-191. In other embodiments of the invention, the peptide comprises a homologue, analogue, mutant, variant or derivative obtained by insertion, deletion or substitution of amino acids in, or chemical modification of, the native carboxyl-terminal sequence of human growth hormone or the growth hormone of a non-human mammalian species.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

According to one aspect of the present invention, there is provided a method for the treatment of obesity in an animal, which comprises administering to the animal an effective amount of a peptide which comprises the carboxyl-terminal sequence of a growth hormone.

Preferably, the animal is a human although the invention also extends to the treatment of non-human mammals. Preferably also, the peptide comprises the carboxyl-terminal sequence of human growth hormone containing amino acid residues 177-191 (hereinafter referred to as hGH 177-191). Alternatively, the peptide may comprise the carboxyl-terminal sequence of the growth hormone of other non-human mammalian species, such as bovine, porcine, ovine or equine growth hormone, corresponding to the hGH 177-191 peptide.

As used throughout this specification, the term "obesity" is used to include both excess body weight and excess adipose tissue mass in the animal, and correspondingly the references to treatment of obesity include both reduction of body weight gain and reduction of adipose tissue mass of the obese animal.

The expected outcome of any treatment of obesity is the reduction of body weight, body adipose tissue mass in particular. The reduction of body adipose tissue mass is directly regulated by two biochemical processes - lipogenesis (fat-production)

and lipolysis (fat-reduction) - and it is generally understood that these biochemical processes are controlled by key metabolic enzymes, specifically the fat-reducing key enzyme (hormone-sensitive lipase) and the fat-producing key enzyme (acetyl CoA carboxylase).

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It has now been shown by the present inventors that hGH 177-191 is effective in stimulating the fat-reducing key enzyme, hormone-sensitive lipase, and in inhibiting the fat-producing key enzyme, acetyl CoA carboxylase. This is further supported by data showing that in the presence of hGH 177-191, fat utilization is accelerated while fat production is reduced, as measured by metabolic end-products *in vitro* as well as *in vivo*. In addition, the mechanism of these molecular actions has been established as resulting from the activation of the production of the cellular second-messenger, diacylglycerol.

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It will, of course, be appreciated that the present invention extends to the use of peptides having longer amino acid sequences than the particular sequence 177-191 of growth hormone, for example the sequence 172-191 of human growth hormone or the corresponding sequence of growth hormone of other non-human mammalian species, however the present invention does not extend to the use of intact full-length human growth hormone or growth hormone of other animal species.

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The present invention also extends to the use of peptides which are homologues, analogues, mutants, variants or derivatives of the native carboxyl-terminal sequences of human growth hormone or growth hormone of other animal species, and which are derived from natural or synthetic (including recombinant) sources, provided always that the resulting peptide retains the biological activity of the native carboxyl-terminal sequence described herein, namely the ability to reduce body weight gain and adipose tissue mass in an obese animal.

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These homologues, analogues, mutants, variants or derivatives may be derived by elongation, insertion, deletion or substitution of amino acids in, chemical modification of, or introduction of a cyclic amide bond between the side chains of amino acids of, the native carboxyl-terminal sequence. Amino acid insertional derivatives include amino and/or carboxylic terminal fusions as well as intra-sequence insertions of single or multiple amino acids. Insertional amino acid sequence variants are those in which one or more amino acid residues are introduced into a predetermined site in the protein although random insertion is also possible with suitable screening of the resulting product. Deletional variants are characterised by the removal of one or more amino acids from the sequence. Substitutional amino acid variants are those in which at least one amino acid residue in the sequence has been replaced by another of the twenty primary protein amino acids, or by a non-protein amino acid. Chemical modifications of the native carboxyl-terminal sequence include the acetylation of the amino-terminus and/or amidation of the carboxyl-terminus and/or side chain cyclisation of the native carboxyl-terminal sequence.

Homologues, analogues, mutants, variants or derivatives of the native carboxyl-terminal sequences of human growth hormone or growth hormone of other animal species which in particular retain the same conformation, structure and charge characteristics as the native carboxyl-terminal sequences can be expected to exhibit the same or similar biological activity as the native sequences, in particular in the ability to reduce body weight gain and adipose tissue mass in an obese animal.

Peptides comprising amino acid residues 177-191 of native human growth hormone (hGH 177-191) include the following sequence:

Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe

(SEQ ID NO: 1)

Such a native peptide may be in cyclic disulfide form, and may comprise an organic or inorganic acid addition salt.

5 Analogues of the hGH 177-191 peptide may be obtained by deletion or insertion of one or more amino acid residues at any position along the native sequence, with the retention of anti-obesity properties as described above.

Alternatively, analogues of the hGH 177-191 peptide may be obtained by substitution of one or more amino acid residues at any position along the native
10 sequence. Such substitution analogues include the following sequence:

AA1-Arg-AA2-Val-Gln-AA3-AA4-Ser-Val-Glu-AA5-AA6-AA3-AA5-AA7

wherein:

(SEQ ID NO: 2)

15 AA1 is selected from the group consisting of L- or D- Leu and Ala
AA2 is selected from the group consisting of L- or D- Ile, Lys and Ala
AA3 is selected from the group consisting of L- or D- Cys and Pen
AA4 is selected from the group consisting of L- or D- Arg and Lys
AA5 is selected from the group consisting of L- or D- Ala and Gly
20 AA6 is selected from the group consisting of L- or D- Ser and Ala
AA7 is selected from the group consisting of L- or D- Phe and Ala

Analogues may also be obtained by elongation of the native hGH 177-191 peptide sequence at one or both ends of the amino acid residues with one or more
25 hydrophilic amino acids to increase solubility in aqueous solution. Such analogues include the following sequence:

Xm-Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe-Xn

(SEQ ID NO: 3)

wherein X is selected from the group consisting of L- or D-Arg, His, Lys and Tyr, and m and n are each 0, 1, 2 or 3 with the provision that at least m or n is 1.

5 Analogues may also be obtained by chemical modification of the native hGH 177-191 peptide sequence. Such analogues include the following sequence:

X-Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe-Y

(SEQ ID NO: 4)

10 wherein X is selected from the group consisting of the desamino form (H), CH₃CO- and other acyl groups; and Y is selected from the group of -COOH, -CONH₂ and alkyl amide groups.

15 Other analogues may also be obtained by side chain cyclization of the native hGH 177-191 peptide sequence, in particular to introduce a cyclic amide bond between amino acid residues 183 and 186 of the native sequence. Such analogues include the following:

Leu-Arg-Ile-Val-Gln-Cys-A-Ser-Val-B-Gly-Ser-Cys-Gly-Phe (SEQ ID NO: 5)

and

20 Leu-Arg-Ile-Val-Gln-Cys-B-Ser-Val-A-Gly-Ser-Cys-Gly-Phe (SEQ ID NO: 6)

wherein A is selected from the group consisting of L- or D- Lys and Orn; and B is selected from the group consisting of L- or D- Glu and Asp.

25 Where appropriate, the analogues described above may also be in cyclic disulfide form, and may comprise an organic or inorganic acid addition salt.

Specific hGH 177-191 analogues and derivatives obtained by substitution of amino acids, elongation of, by chemical modification of, or by introduction of a cyclic

amide bond between side chains of amino acids, of the native hGH 177-191 peptide sequence, and which exhibit anti-obesity properties, include the following:

- 5 (i) Lys-Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe (SEQ ID NO: 7)
- (ii) Lys-Lys-Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe (SEQ ID NO: 8)
- (iii) Ala-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe (SEQ ID NO: 9)
- 10 (iv) Leu-Arg-Ala-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe (SEQ ID NO: 10)
- (v) Leu-Arg-Lys-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe (SEQ ID NO: 11)
- (vi) Leu-Arg-Ile-Val-Gln-D-Cys-Arg-Ser-Val-Glu-Gly-Ser-D-Cys-Gly-Phe (SEQ ID NO: 12)
- 15 (vii) Leu-Arg-Ile-Val-Gln-Pen-Arg-Ser-Val-Glu-Gly-Ser-Pen-Gly-Phe (SEQ ID NO: 13)
- (viii) Leu-Arg-Ile-Val-Gln-D-Pen-Arg-Ser-Val-Glu-Gly-Ser-D-Pen-Gly-Phe (SEQ ID NO: 14)
- 20 (ix) Leu-Arg-Ile-Val-Gln-Cys-D-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe (SEQ ID NO: 15)
- (x) Leu-Arg-Ile-Val-Gln-Cys-Lys-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe (SEQ ID NO: 16)
- (xi) Leu-Arg-Ile-Val-Gln-Cys-D-Lys-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe (SEQ ID NO: 17)
- 25 (xii) Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Ala-Ser-Cys-Ala-Phe (SEQ ID NO: 18)
- (xiii) Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-D-Ala-Ser-Cys-D-Ala-Phe (SEQ ID NO: 19)
- 30 (xiv) Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ala-Cys-Gly-Phe (SEQ ID NO: 20)
- (xv) Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Ala-Phe (SEQ ID NO: 21)
- (xvi) Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Ala (SEQ ID NO: 22)
- 35 (xvii) Leu-Arg-Ile-Val-Gln-Cys-Lys-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe (SEQ ID NO: 23)
- (xviii) Leu-Arg-Ile-Val-Gln-Cys-Orn-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe (SEQ ID NO: 24)
- 40 (xix) Leu-Arg-Ile-Val-Gln-Cys-Lys-Ser-Val-Asp-Gly-Ser-Cys-Gly-Phe (SEQ ID NO: 25)
- (xx) Leu-Arg-Ile-Val-Gln-Cys-Orn-Ser-Val-Asp-Gly-Ser-Cys-Gly-Phe (SEQ ID NO: 26)
- (xxi) Leu-Arg-Ile-Val-Gln-Cys-Glu-Ser-Val-Lys-Gly-Ser-Cys-Gly-Phe

- (SEQ ID NO: 27)
 (xxii) Leu-Arg-Ile-Val-Gln-Cys-**Glu**-Ser-Val-**Orn**-Gly-Ser-Cys-Gly-Phe
 (SEQ ID NO: 28)
 (xxiii) Leu-Arg-Ile-Val-Gln-Cys-**Asp**-Ser-Val-**Lys**-Gly-Ser-Cys-Gly-Phe
 (SEQ ID NO: 29)
 (xxiv) Leu-Arg-Ile-Val-Gln-Cys-**Asp**-Ser-Val-**Orn**-Gly-Ser-Cys-Gly-Phe
 (SEQ ID NO: 30)
 (xxv) desaminoLeu-hGH 177-191 (SEQ ID NO: 31)
 (xxvi) acetyl-hGH 177-191 (SEQ ID NO: 32)
 (xxvii) hGH 177-191-amide (SEQ ID NO: 33)

wherein the amino acid residue abbreviations used are in accordance with the standard peptide nomenclature:

15	Gly	=	Glycine;	Ile	=	Isoleucine;
	Glu	=	Glutamic Acid;	Phe	=	Phenylalanine;
	Cys	=	Cysteine;	Arg	=	Arginine;
	Gln	=	Glutamine;	Leu	=	Leucine;
	Ser	=	Serine;	Val	=	Valine;
20	Lys	=	Lysine;	DAla	=	Alanine;
	Asp	=	Aspartic acid;	His	=	Histidine;
	Orn	=	Ornithine;	Tyr	=	Tyrosine;
	Pen	=	Pencillamine (β,β' -Dimethyl-L-Cysteine).			

25 All amino acids, except for glycine, are of the L-absolute configuration, unless indicated as D-absolute configuration.

Where appropriate, the analogues described above may be in cyclic disulfide form, and may comprise an organic or inorganic acid addition salt. In the case of
 30 analogues SEQ ID NO: 23 to SEQ ID NO: 30. there may be a cyclic amide bond between amino acid residues in bold.

The term "effective amount" as used herein means an amount of the peptide sufficient to attain the desired effect in the treatment of obesity in the animal, but not so large an amount as to cause serious side effects or adverse reactions.

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In another aspect, the present invention provides the use of an effective amount of a peptide which comprises the carboxyl-terminal sequence of a growth hormone as described above, or a homologue, analogue, mutant, variant or derivative thereof as described above, in the treatment of obesity in an animal or
10 in the manufacture of a pharmaceutical composition for the treatment of obesity in an animal.

In yet another aspect, the present invention provides a pharmaceutical composition for use in the treatment of obesity in an animal, comprising an effective
15 amount of a peptide which comprises the carboxyl-terminal sequence of a growth hormone as described above, or a homologue, analogue, mutant, variant or derivative thereof as described above, together with one or more pharmaceutically acceptable carriers and/or diluents.

20 The peptide which is the active ingredient of the pharmaceutical composition of this aspect of the invention exhibits advantageous therapeutic activity in the treatment of obesity in an animal when administered in an amount appropriate to the particular case. For example, from about 0.5 μ g to about 20 mg per kilogram of body weight per day may be administered. Dosage regimens may
25 be adjusted to provide the optimum prophylactic or therapeutic response. For example, one or more divided doses may be administered daily, weekly, monthly or in other suitable time intervals or the dose may be proportionally reduced as indicated by the exigencies of the clinical situation.

The active ingredient may be administered in any convenient manner such as by the oral, parenteral (including intraperitoneal, intravenous, subcutaneous, intramuscular and intramedullary injection), intranasal, intradermal or suppository routes or by implanting (eg using slow release devices). For ease of administration, oral administration is preferred, however parenteral administration is also quite convenient. Depending on the route of administration, the active ingredient may be required to be coated in a material that protects said ingredient from the action of enzymes, acids and other natural conditions which may inactivate the said ingredient. For example, low lipophilicity of the ingredient may allow it to be destroyed in the gastrointestinal tract by enzymes capable of cleaving peptide bonds and in the stomach by acid hydrolysis. In order to administer the composition by other than parenteral administration, the active ingredient may be coated by, or administered with, a material to prevent its inactivation.

The active ingredient may also be administered in dispersions prepared in glycerol, liquid polyethylene glycols, and/or mixtures thereof and in oils. Under ordinary conditions of storage and use, these preparations will usually contain a preservative to prevent the growth of microorganisms.

The pharmaceutical forms suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. In all cases, the form must be sterile and must be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and vegetable oils. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion

- 13 -

and by the use of surfactants. The prevention of the action of microorganisms can be brought about by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thiomorosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by, for example, the use in the compositions of agents delaying absorption.

Sterile injectable solutions are prepared by incorporating the active ingredient in the required amount in the appropriate solvent with various of the other ingredients enumerated above, as required, followed by filtered sterilisation. Generally, dispersions are prepared by incorporating the sterilised active ingredient into a sterile vehicle which contains the basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum drying and the freeze-drying technique which yield a powder of the active ingredient plus any additional desired ingredient from previously sterile-filtered solution thereof.

When the active ingredient is suitably protected as described above, the composition may be orally administered, for example, with an inert diluent or with an assimilable edible carrier, or it may be enclosed in hard or soft shell gelatin capsule, or it may be compressed into tablets, or it may be incorporated directly with the food of the diet. For oral administration, the active ingredient may be incorporated with excipients and used in the form of ingestible tablets, buccal tablets, troches, capsules, elixirs, suspensions, syrups, wafers, and the like. Such compositions and preparations should contain at least 0.01% by weight and more preferably at least 0.1-1% by weight of active ingredient. The percentage of the compositions and preparations may, of course, be varied and may conveniently be between about 5 to about 80% of the weight of the unit. The amount of active ingredient in the pharmaceutical compositions is such that a suitable dosage will be

obtained. Preferred compositions or preparations according to the present invention may, for example, be prepared so that an oral dosage unit form contains between about 0.5 µg and 200 mg and more preferably 10µg and 20 mg of active ingredient.

5 The tablets, troches, pills, capsules and the like may also contain the following: a binder such as gum tragacanth, acacia, corn starch or gelatin; excipients such as dicalcium phosphate; a disintegrating agent such as corn starch, potato starch, alginic acid and the like; a lubricant such as magnesium stearate; and a sweetening agent such a sucrose, lactose or saccharin may be added or a
10 flavouring agent such as peppermint, oil of wintergreen, or cherry flavouring. When the dosage unit form is a capsule, it may contain, in addition to materials of the above type, a liquid carrier. Various other materials may be present as coatings or to otherwise modify the physical form of the dosage unit. For instance, tablets, pills, or capsules may be coated with shellac, sugar or both. A syrup or elixir may contain
15 the active compound, sucrose as a sweetening agent, methyl and propylparabens as preservatives, a dye and flavouring such as cherry or orange flavour. Of course, any material used in preparing any dosage unit form should be pharmaceutically pure and substantially non-toxic in the amounts employed. In addition, the active ingredient may be incorporated into sustained-release preparations and
20 formulations.

 As used herein, pharmaceutically acceptable carriers and diluents include any and all solvents, dispersion media, aqueous solutions, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like. The
25 use of such media and agents for pharmaceutically active substances is well known in the art, and it is described, by way of example, in *Remington's Pharmaceutical Sciences*, 18th Edition, Mack Publishing Company, Pennsylvania, USA. Except insofar as any conventional media or agent is incompatible with the active ingredient, use thereof in the pharmaceutical compositions of the present invention

is contemplated. Supplementary active ingredients can also be incorporated into the compositions.

5 It is especially advantageous to formulate compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used herein refers to physically discrete units suited as unitary dosages for the human subjects to be treated; each unit containing a predetermined quantity of active ingredient calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier and/or diluent. The specifications for the novel
10 dosage unit forms of the invention are dictated by and directly dependent on (a) the unique characteristics of the active ingredient and the particular therapeutic effect to be achieved, and (b) the limitations inherent in the art of compounding such an active ingredient for the treatment of obesity.

15 Throughout this specification and claims which follow, unless the context requires otherwise, the word "comprise", or variations such as "comprises" or "comprising", will be understood to imply the inclusion of a stated integer or group of integers but not the exclusion of any other integer or group of integers.

20 Further details of the present invention will be apparent from the following Example and the accompanying drawings which are included by way of illustration, not by way of limitation, of this invention.

BRIEF DESCRIPTION OF THE DRAWINGS.

25

In the drawings:

Figures 1A & 1B show the effect of hGH 177-191 peptide on cumulative body weight gains in male (1A) and female (1B) C57BL/6J (ob/ob) mic during the
30 18-day treatment period. Animals were given a daily intraperitoneal injection of 0.1

ml of either saline or hGH 177-191 (200 μ g/kg body weight). Each points represents the mean \pm SEM of 6 animals.

Figures 2A & 2B show the average daily food consumption (g/mouse/day) of C57BL/6J (ob/ob) mice during an 18-day treatment period with hGH 177-191. The treatment for the four groups of animals was as described in figures 1A & 1B. Each point represents the mean \pm SEM of 6 animals. No significance between the groups was observed at all times.

Figures 3A & 3B demonstrate the effect of hGH 177-191 peptide on body weight gain of 14-15 week old male Zucker fatty (fa/fa) rats during the 20 or 27-days treatment period. Animals were given a daily intraperitoneal injection of either saline or the peptide (500 μ g/kg body weight) (3A) or implanted intradermally with a slow-release tablet (500 μ g/day/kg body weight) in the lower quadrant of the abdomen of the animals. The control group was implanted with a placebo tablet in the same manner. Each point represents the mean \pm SEM of 6 animals.

Figures 4A & 4B demonstrate the average daily food consumption (g/rat/day) of Zucker fatty rats during the treatment period with hGH 177-191 peptide. The treatment for the four groups of animals was as described in Figures 3A & 3B. Each point represents the mean \pm SEM of 6 animals. No significance between the test groups and the appropriate controls was observed at all times

Figures 5A & 5B depict the *ex vivo* effect on lipogenesis in adipose tissues of the C57BL/6J (ob/ob) male mice (5A) and female mice (5B) after 18-day treatment with hGH 177-191. Data indicate the rate of [C^{14}]-lipid synthesis and are expressed as [C^{14}]-glucose incorporated into lipid (pmol/mg tissue/min) Values are mean \pm SEM of 12 determinations from 6 animals of each group. The differences between the hGH 177-191 treated and saline control groups were statistically significant.

Figure 6A & 6B depict the *ex vivo* effect on lipolysis in adipose tissue of the C57BL/6J (ob/ob) male mice (6A) and female mice (6B) after 18-days treatment with hGH 177-191. Data indicate the rate of glycerol release from adipose tissues (pmol/mg tissue/min). Values are mean \pm SEM of 12 determinations from 6 animals of each group. The differences between the hGH 177-191 treated and saline control groups were statistically significant.

Figure 7 illustrates the *in vitro* effect of hGH 177-191 on fatty acid oxidation in isolated adipose tissues of C57BL/6J (ob/ob) mice with the determination of the rate of $[C^{14}]O_2$ production from $[C^{14}]$ -palmitic acid. The rate of $[C^{14}]$ -palmitic acid oxidation was expressed as $\mu\text{mol/g tissue/hr}$.

Figure 8 illustrates the *in vitro* effect of hGH 177-191 on the release of diacylglycerol from isolated adipocytes of normal rats over an incubation period of 40 min. Diacylglycerol was quantitated using radioenzymic assay and the results were expressed as % increase over the basal levels.

Figure 9 shows the *in vitro* effect of hGH 177-191 on hormone-sensitive lipase activity in isolated adipocytes of male Zucker fatty (fa/fa) rats was determined by the amount of hydrolyzed $[C^{14}]$ -oleic acid from $[C^{14}]$ -triolein. The enzyme was expressed as U/mg protein, where the release of 1 nmole of oleic acid per hour was considered as 1 Unit of enzyme activity.

Figures 10A & 10B shows the *in vitro* effect of hGH 177-191 on acetyl-CoA carboxylase in the isolated adipocytes (10A) and hepatocytes (10B) of normal rats was determined by $[C^{14}]$ -bicarbonate fixation reaction and expressed as mU/g cell dry weight, where 1 Unit of acetyl-CoA carboxylase was defined as the carboxylation of 1 μmole acetyl-CoA per minute.

Figure 11A demonstrates the effect of an analogue SEQ ID NO: 11 on body weight gain of 26-week-old C57BL/6J (ob/ob) mice during the 18 days treatment period. Animals were given a daily intraperitoneal injection of either saline (as control) or peptide analogue (500 µg/kg body weight). Each point represents the mean ± SEM of 6 animals.

Figure 11B shows the average daily food consumption (g/mouse/day) of 26-week-old C57BL/6J (ob/ob) mice during the treatment period with analogue SEQ ID NO: 11. The treatment for the two groups of animals was as described in Figure 11A. Each point represents the mean ± SEM of 6 animals. No significance between the test group and the control was observed at all times.

EXAMPLE

MATERIALS AND METHODS

Animals and treatments.

Obese C57BL/6J (ob/ob) mice and fatty Zucker (fa/fa) rats were used to demonstrate the biological effects of the synthetic hGH 177-191 and analogues. The animals of the same age and same sex were randomly divided into two groups, housed 6 per cage and maintained on a normal 12-hr light/dark cycle at a constant room temperature of 25°C in the animal house of the Department of Biochemistry and Molecular Biology, Monash University, Clayton, Australia. Animals were fed *ad libitum* on pre-determined quantity of animal pellets (Clark King, Melbourne, Australia) and allowed free access to water at all times. The animals were given a daily intraperitoneal (i.p) injection of 0.1 ml of either the synthetic peptide (200-500µg/kg body weight) or equivalent volume of physiological saline (0.9% sodium chloride) for appropriate number of days. The i.p. injection was administered with a 30G x ½" (0.31 x 13 mm) needle on a 1-ml disposable tuberculin syringes, and the site of injection was the lower left quadrant of the

abdomen of the animals. To study the effects of controlled-delivery of hGH 177-191 and analogues, slow-release peptide-pellets in a diameter of 3 mm were implanted intradermally in the abdominal region of Zucker rats under anaesthesia. The body weight and food intake were monitored for the periods of time as indicated.

5

Peptide Synthesis

The peptides of the present invention were prepared by using standard 9-fluorenylmethyloxycarbonyl(Fmoc) solid-phase techniques. The solid-phase synthesis, for example, could be commenced from the C-terminal end of the peptide using an α -amino protected amino acid. A suitable starting materials could be prepared, for instance, by attaching the required α -amino acid to a Wang resin (4-alkoxybenzyl alcohol resin), or Rink amide resin (2,4-dimethoxy-4'-[carboxymethoxy]-benzhydrylamine linked to amino methyl resin) or PAM resin (4-hydroxymethylphenyl- acetic acid resin). Resins were commercially available from Auspep Pty. Ltd., Parkville, Victoria, Australia.

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In the solid-phase preparation of the compounds of this invention, a protected amino acid was coupled to a resin with the aid of a coupling agent. After the initial coupling, the α -amino protecting group was removed by piperidine in organic solvents at room temperature. Following the removal of the α -amino protecting group, the remaining protected amino acids were coupled stepwise in the desired order. A 4-fold excess of each protected amino acid was generally used in the reaction with an appropriate carboxyl group activator such as diisopropylcarbodiimide (DIC) and 1-hydroxybenzotriazole (HOBt), in methylene chloride (DCM)-N,N-dimethylformamide (DMF) mixtures.

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After the desired amino acid sequence was completed, the peptide was then cleaved from the resin support by treatment with a reagent such as trifluoroacetic acid (TFA) or trifluormethanesulfonic acid (TFMSA) which cleaved the peptide from the resin, as well as all side-chain protecting groups, except for

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Cys(Acm). When a Wang resin was used, TFA treatment resulted in the formation of the free peptide acids. When the Rink amide resin was used, TFA treatment resulted in the formation of the free peptide amides. When a PAM resin was used, TFMSA treatment resulted in the formation of the free peptide acids. The target peptides might exist in cyclic disulfide form which requires post-synthesis modification.

The following examples are provided for the purpose of further illustration only and not intended to be limitations on the disclosed invention.

A. Synthesis of pentadecapeptide comprising amino acid residues 177-191 of native human growth hormone, designated as hGH (177-191):

Leu¹-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe¹⁵ (cyclic disulfide)

The following procedure was employed in preparing the pentadecapeptide:

Step 1. Wang resin (0.625 g, 0.5 mmol) was placed in a 10 ml-reaction vessel. DCM (4 ml) was added to the reaction vessel. The Wang resin was washed with vigorous stirring for 2 minutes. The DCM solution was then drained from the reaction vessel. This washing was repeated twice.

Step 2. Fmoc-L-phenylalanine (Fmoc-Phe, 0.388 g, 1.0 mmol) in 2.4 ml NMP-DCM (1:5, v/v) and DIC (0.135 g, 1.0 mmol) in 1.0 ml NMP were mixed in a reaction vessel for 10 minutes. To the mixture, 4-dimethylaminopyridine (DMAP, 0.074g, 0.06 mmol) in 0.6 ml DMF was added. The reaction in the solution was allowed to continue for 68 minutes at room temperature. The solution was then drained, the resin was washed thoroughly with NMP (4 ml x 3) and DCM (4 ml x 3). The Fmoc-Phe-Wang Resin complex was dried *in vacuo* overnight to yield 0.781 g of material. The coupling level of amino acid to resin was determined to be 0.80 mmol/g resin by using spectrophotometric measurement of the Fmoc-piperidine adduct.

Step 3. Fmoc-Phe-Wang Resin (0.263 g, 0.20 mmol) was placed in the 10 ml-reaction vessel. DMF (8 ml) was added to wash and swell the resin by stirring for 2 minutes. The solution was then drained from the reaction vessel.

Step 4. A 25% piperidine/DMF solution (4 ml) was added to the reaction vessel. The resulting mixture was stirred for 2 minutes. The solution was drained from the reaction vessel. This deprotection procedure was repeated once but with prolonged stirring time (18 min). The solution was drained from the reaction vessel.

Step 5. DMF (8 ml) was added to the reaction vessel. The resulting solution was stirred for 2 minutes. The solution was drained from the resin in the reaction vessel. This washing procedure was repeated twice. DMF (2 ml) was added to the reaction vessel to keep the resin swollen.

Step 6. Fmoc-glycine (Fmoc-Gly, 0.238 g, 0.8 mmol), HOBt (108 mg; 0.8 mmol) and DIC (128 μ l; 0.8 mmol) were added to a 10 ml-test tube containing 2 ml DMF. The mixture was stirred for 10 minutes to start the activation of amino acid. The solution was then added to the resin which originally was placed in the reaction vessel. The resulting mixture was stirred for 1.5 hours or until a negative ninhydrin test was obtained. The solution was then drained from the reaction vessel.

Step 7. DMF (8 ml) was added to the reaction vessel. The resulting solution was vigorously stirred for 2 minutes. The solution was then drained from the reaction vessel. The washing procedure was repeated twice.

Steps 4 through 7 were then repeated employing the following order of amino acid:

Fmoc-Cys(Acm)
Fmoc-Ser(t-Bu)
Fmoc-Gly
Fmoc-Glu(O-tBu)
Fmoc-Val

Fmoc-Ser(t-Bu)

Fmoc-Arg(Pmc)

Fmoc-Cys(Acm)

Fmoc-Gln

5 Fmoc-Val

Fmoc-Ile

Fmoc-Arg(Pmc)

Fmoc-Leu

10 After completion of the synthesis of the desired peptide-resin, the reaction vessel containing the peptide-resin was then placed in a desiccator and dried overnight under vacuum. The yield of peptide-resin was 0.635 g. The dried peptide-resin was removed from the reaction vessel and placed in a 50 ml round-bottom flask containing a magnetic stirring bar. The cleavage of the peptide from the resin
15 with TFA was carried out with the following procedure: A scavenger solution, containing 0.75 g phenol, 0.5 ml H₂O, 0.5 ml thioanisole, and 0.25 ml ethanedithio, was added to the round-bottom flask. The resulting mixture was stirred for 5 minutes. 10 ml TFA was added drop by drop into the flask while kept stirring vigorously. The resulting mixture was stirred for 2.5 hours at room temperature.

20

The mixture was filtered through a medium-porosity filter, fritted glass funnel. The TFA-peptide solution was sucked into another 500 ml round-bottom flask containing 200 mL cold diethyl ether by applying vacuum. Peptide was allowed to be precipitated in the ether solution at 4°C overnight, then collected by filtering the
25 mixture through a fine-porosity, fritted glass funnel. The peptide pellet on the filter was washed with cold ether (10 ml x 3) to remove the scavenger. The peptide pellet was then dissolved with 25% aqueous acetic acid and then lyophilized to yield the crude peptide (about 400 mg dry weight, purity ~80%).

The crude peptide was purified by reversed-phase high performance liquid chromatography (RP-HPLC). Purification was carried out on a preparative 21.2 x 250 mm Supelcosil PLC-18 (octadecyl, C₁₈) column (120 Angstrom pore size, 12 µm particle size, 190 m²/g surface area; Supelco, Bellefonte, PA, U.S.A.) at 5.0 ml/min flow rate at room temperature. A linear gradient program was utilized, where solvent A was water with 0.1% TFA, and solvent B was acetonitrile-water (50/50: v/v, containing 0.1% TFA). The gradient was developed from 20 to 100% over 80 min. Separation profiles were recorded and analysed using a Perkin-Elmer LC-100 integrator. The desired peptide component was eluted and collected with the Pharmacia Model FRAC-100 automatic fraction collector (Uppsala, Sweden). The fractions of identical component were combined and lyophilized. The purified peptide (275 mg dry weight, purity >98%), Cys(Acm)^{6,13}-pentadecapeptide, was kept frozen at -20°C.

For cyclisation of the disulfide bridge of the peptides, iodine oxidation in 80% aqueous acetic acid was used to remove the cysteine-protecting groups, Acm, and furnished the intramolecular disulfide bridge simultaneously. Cys(Acm)^{6,13}-pentadecapeptide (275 mg, 0.155 mmole) was dissolved in 50 ml 80% aqueous acetic acid. This solution was slowly added to a 250 ml round-bottom flask containing iodine (378 mg, 1.4 mmole) in 100 ml 80% aqueous acetic acid by stirring vigorously. Reaction was allowed to continue for 2 hours at room temperature and terminated by adding the ascorbic acid (Vitamin C) to the resulting solution. Liquid volume was then reduced by rotary evaporation and peptide recovered by lyophilization. The cyclised peptide was then purified by RP-HPLC as described in the purification of linear peptide. After lyophilization, 165 mg cyclic pentadecapeptide with 96% purity was yielded. The total yield of synthesis was about 46%.

B. Synthesis of pentadecapeptide:

Leu¹-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe¹⁵-amide
(cyclic disulfide)

The procedure set forth in EXAMPLE A was employed. The modification
5 consisted of omitting Wang resin and replacing it with the Rink amide resin.

C. Synthesis of the pentadecapeptide:

desamino-Leu¹-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe¹⁵
10 (cyclic disulfide)

The procedure set forth in EXAMPLE A was employed. The modification
consisted of the replacement of Fmoc-Leu and with 4-methyl-pentacarboxylic acid,
resulting in the synthesis of the desamino pentadecapeptide.

15

D. Synthesis of the pentadecapeptide:

CH₃CO-Leu¹-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe¹⁵
(cyclic disulfide)

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The procedure set forth in EXAMPLE A was employed. After completion of the
synthesis of the desired deblocked peptide-resin, 5 ml solution of 20% acetic
anhydride in DMF was added. After 5 minutes, 71 μ l (0.4 mmols) of
diisopropylethylamine (DIEA) was added to neutralize the protons that were
25 generated. Acetylation of the peptide was performed at room temperature for 30
minutes. The peptide-resin was washed twice with DMF and twice with DCM and
the N-acetyl peptide resin was ready for TFA cleavage as shown in EXAMPLE A.

E. Synthesis of the dicyclo-pentadecapeptide:

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Leu¹-Arg-Ile-Val-Gln-Cys-Lys-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe¹⁵

(cyclic disulfide)

The following procedure was employed in preparing the pentadecapeptide:

- 5 **Step 1.** Boc-L-phenylalanine-PAM resin (0.400 g, 0.2 mmol; Auspep, Melbourne, Australia; Cat#5290F, Batch#494123) was placed in a 10ml reaction vessel. The resin was washed with DCM (4 ml) by vigorous stirring for 2 minutes. The DCM solution was then drained from the reaction vessel. This washing was repeated once.
- 10 **Step 2.** 50% TFA/DCM solution (4 ml) was added to the reaction vessel. The resulting mixture was stirred for 2 minutes. The solution was then drained from the reaction vessel. This deprotection procedure was repeated once with a stirring time of 18 minutes. The solution was drained from the reaction vessel. DCM (4 ml) was added to the reaction vessel and the content was allowed
- 15 to stand for 2 minutes. The solution was again drained from the resin. This washing procedure was repeated twice. 10% DIEA/DMF (4ml) was added to the reaction vessel. The resulting mixture was allowed to stand for 1 minute and the solution removed as before. This deprotection procedure was repeated once. DMF (4ml)
- 20 was added to the resin complex in the reaction vessel. The resulting solution was allowed to stand for 2 minutes, followed by the removal of the solution from the vessel. This washing procedure was repeated four times. Finally, DMF (2 ml) was added to the reaction vessel to keep the resin swollen.
- 25 **Step 3.** Fmoc-glycine (Fmoc-Gly, 0.238 g, 0.8 mmol), HOBt (108 mg; 0.8 mmol) and DIC (128 μ l; 0.8 mmol) were added to a 10 ml-test tube containing 2 ml DMF. The mixture was stirred for 10 minutes to activate the amino acid. The solution was then added to the resin in the reaction vessel. The resulting mixture was stirred for 1.5 hours or until a negative ninhydrin test was obtained. The solution was then drained from the reaction vessel.

Step 4. DMF (8 ml) was added to the reaction vessel. The resulting solution was vigorously stirred for 2 minutes, followed by the removal of the supernatant. The washing procedure was repeated twice.

5 **Step 5.** 25% piperidine/DMF solution (4 ml) was added to the reaction vessel. The resulting mixture was stirred for 2 minutes. The solution was drained from the reaction vessel. This deprotection procedure was repeated once with stirring for 18 minutes. The solution was drained from the reaction vessel.

10 **Step 6.** DMF (8 ml) was added to the reaction vessel. The resulting solution was stirred for 2 minutes. The solution was drained from the reaction vessel resin. This washing procedure was repeated twice. 2 ml DMF was added to the reaction vessel to keep the resin swollen.

Steps 3 through 6 were then repeated with the following order of amino acids:

15 Fmoc-Cys(Acm)
Fmoc-Ser(Bzl)
Fmoc-Gly
Fmoc-Glu(O-tBu)
Fmoc-Val
Fmoc-Ser(Bzl)

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Step 7. Steps 3 and 4 were repeated to couple Fmoc-Lys(Boc) to Ser in the position 184. After completion of coupling, the reaction vessel containing the peptide-resin was then placed in a desiccator and dried overnight under vacuum. the peptide-resin was then transferred to a 10ml-reaction vessel. DCM (4 ml) was added to the reaction vessel. The resin was washed with vigorous stirring for 2 minutes. The DCM solution was then drained from the reaction vessel. This washing was repeated once.

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Step 8. Step 2 was used to remove the Boc group and the t-Bu group from the side chain of lysine and glutamic acid, respectively.

Step 9 1 ml of 1.5% DIEA/DMF was added to the reaction vessel. Benzotriazo-1-yl-oxy-tris-(dimethylamino)phosphonium hexafluorophosphate (BOP) (400 mg; 0.90 mmol), HOBt (122 mg, 0.90 mmol), and DIEA (400 μ l, 2.25 mmol) were dissolved in 3.4 ml of 1.5% DIEA/DMF and then added to the reaction vessel.

5 The resulting mixture was stirred for 3 hours or until a negative ninhydrin test was obtained before the removal of the supernatant from the reaction vessel. DMF (8 ml) was then added to the reaction vessel. The resulting solution was vigorously stirred for 2 minutes. The solution was then drained from the reaction vessel. The washing procedure was repeated twice.

10 **Step 10.** Steps 5 through 6 were then repeated to remove the Fmoc group from the α -amino group of the lysine residue in the peptide-resin.

Steps 3 through 6 were then repeated with the following order of amino acids:

15 Fmoc-Cys(Acm)
Fmoc-Gln
Fmoc-Val
Fmoc-Ile
Fmoc-Arg(Pmc)
Fmoc-Leu

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After completion of the synthesis of the desired peptide resin, Step 4 was then repeated to remove the Fmoc group from Leu and get the deblocked peptide resin. The reaction vessel containing the peptide resin was then placed in a desiccator and dried overnight under vacuum. 604 mg peptide-resin was yielded. The dried peptide resin was removed from the reaction vessel and placed in a 25 ml round-bottom flask containing a magnetic stirring bar. Trifluoromethanesulfonic acid (TFMSA)/TFA cleavage protocol was used to cleave peptide from the PAM resin: A scavenger solution, containing 500 μ l thioanisole, and 250 μ l ethanedithio, was added to the flask. The resulting mixture was stirred for 10 minutes at room temperature. 5 ml of TFA was added drop by drop into the flask while kept stirring

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vigorously. The resulting mixture was stirred at room temperature for 15 minutes. Place the flask in an ice-bath and then 500 ml of TFMSA was slowly added while kept stirring vigorously. The resulting mixture was stirred in the ice-bath for 10 minutes and at room temperature another 15 minutes. Cold diethyl ether (50 ml) was added to the flask to stop the reaction and precipitate the cleaved peptide. The peptide was collected by filtering the mixture through a fine-porosity, fritted glass funnel and washed with cold ether (10 mL x 3) to remove the scavenger. The peptide pellet was dissolved with 30 ml of 50 % acetonitrile/H₂O followed by the addition of 5 ml of cold 10% NH₄HCO₃ to neutralize the solution. The crude peptide (519 mg, purity ~69%) was obtained after lyophilization.

The preparation of peptide with cyclic disulfide form and the purification of pure final product are shown in EXAMPLE A.

15 Cumulative weight gain and food consumption.

Cumulative weight gain and food consumption were determined at 3-day intervals by the measurements of body weight and uneaten food remaining in the cages. The animals were placed in a covered chamber to minimise movement during the weighing procedure. The food consumption data were obtained by subtracting the amount of uneaten food remaining in the cages from the original provision.

Assays for plasma triglyceride and total cholesterol.

The animals were anaesthetised with sodium pentobarbitone (80 mg/kg body weight) 12 hr after the last dose of hGH 177-191. Blood samples were collected from the tail vein of anaesthetised animals 45 mins after the administration of anaesthetic. After being centrifuged at 2000 x g for 5 minutes, plasma was removed from the samples and used for metabolite assays. Triglyceride and total cholesterol in plasma were measured by enzyme-spectrophotometric methods. The reagents are based on either a modified glycerol phosphate oxidase (GPO)-

Trinder's type colour reaction²⁴ or a cholesterol oxidase-4-aminoantipyrine method²⁵. All assays were performed with the CentrifChem System 400 (Union Carbide) containing an automated pipetter, a centrifugal analyser and a recording spectrophotometer. Seronorm Lipid (Nycomed Pharma Co., Oslo, Norway) was used as the calibrator.

Determination of adipose tissue weight.

The procedure for the isolation and measurement of intact epididymal fat pads was established in previous studies of epididymal growth of GH-deficient (lit/lit) mice. In the present study, white adipose tissues, either whole epididymal or parametrical fat pads, were excised with the identical techniques as previously described²² from the mice immediately after sacrifice. The tissues were washed in cold physiological saline, blotted and weighed. For *ex vivo* lipogenic assays, the portions of adipose tissues without blood vessels were used.

Hormone-sensitive lipase (HSL) Assay

Hormone-sensitive lipase (HSL) activity of isolated adipocytes was used as a model to evaluate the lipolytic effect of hGH 177-191 peptide and analogues. In this assay [¹⁴C]-triolein was used as a substrate by HSL. The amount of hydrolyzed [¹⁴C]-oleic acid was determined and used as an index of HSL activity.

The adipocytes were prepared from the epididymal fat pads of male Zucker fatty (fa/fa) rats by collagenase digestion. Fat pads (5 g) were finely-cut into small (2 -3 mm) pieces and placed in a siliconised glass vial containing 10 ml digestion medium. The digestion medium contained microbial collagenase (Type II) at a concentration of 1 mg/ml in Krebs-Ringer phosphate buffer (pH 7.4) at half Ca²⁺ strength, at 2% (w/v) of bovine serum albumin (BSA Fraction V). After digestion at 37°C for 1 hr under an atmosphere of 95% O₂/5% CO₂, adipocytes were liberated from any remaining pieces of adipose tissue by gently sucking the suspension up and down with a 5.0 ml pipette with the tip opening of 3 - 4 mm. The adipocytes

released from tissue were then filtered through nylon chiffon into a siliconised glass tube and washed twice with 6 ml of collagen free albumin buffer. The isolated adipocytes were resuspended with 10 ml collagen-free buffer and the concentration of adipocytes was estimated by counting an aliquot of a pre-determined volume of cells on a microscope slide. It normally gave approximate 10^9 cells/ml of adipocytes in Krebs-Ringer phosphate buffer (pH 7.4).

The HSL activity was measured at 37 °C for 1 hour in a final volume of 200 μ l, containing 10 μ moles phosphate buffer (pH 7.0), 15 μ moles of emulsified [C^{14}]-triolein, and 10^8 cells. The substrate, [C^{14}]-triolein, was pre-emulsified with unlabelled triolein to provide the final emulsion contains 15 μ moles of triolein and 375,000 cpm in 0.1 ml.. Different concentrations of hGH 177-191 peptide or analogues were added to evaluate their effects on the HSL activities. The reaction was stopped by adding 1 ml of the fatty acid extraction mixture of choloform-methanol-benzene 2:2:4:1 containing 50 μ g oleic acid, followed by adding 67 μ l of 0.5N NaOH. To extract and isolate free fatty acid, samples were vortexed for 20 seconds and then centrifuged at 1,000 x g for 5 minutes. A 200 μ l portion of the alkaline aqueous upper phase containing fatty acids was transferred to scitillation vials. The [C^{14}]-radioactivity was measured by a liquid scintillation counter. The remaining cell suspension was assayed for protein content.. The HSL activity was expressed as U/mg protein, where the release of 1 nmole of oleic acid per hour was defined as 1 U of enzyme activity.

Acetyl-CoA Carboxylase Assay

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Acetyl-CoA carboxylase catalyzes the critical step in fatty acid synthesis. The acetyl-CoA carboxylase activities of both isolated adipocytes and hepatocytes in the presence of hGH 177-191 peptide or analogues were measured for the evaluation of the anti-lipogenic effect of the peptides. The acetyl-CoA carboxylase activity was

determined by the $[C^{14}]$ -bicarbonate fixation reaction - the rate of incorporation of acetyl-CoA dependent $H[C^{14}]O_3$ into $[C^{14}]$ -malonyl-CoA.

Adipocytes were prepared with the method described in the HSL assay.

5 Hepatocytes were prepared from the livers of male Wistar rats by collagenase digestion. The liver was finely-cut with scissors and transferred to a 250 ml Erlenmeyer flask containing 30 ml of digestion medium. The digestion medium contained microbial collagenase (Type IV) at a concentration of 30 mg/ml in calcium-free Krebs-Ringer phosphate buffer (pH 7.4) and glucose (5 mM). After

10 digestion for 15 minutes at 37°C under an atmosphere of 95% O_2 /5% CO_2 the hepatocytes liberated from the tissue were then filtered through nylon chiffon into a siliconised glass tube and washed twice with fresh collagen-free digestion medium. The isolated cells were resuspended in 45 ml of medium containing extra EDTA (0.45 mmoles), gelatine (0.7 ml), 2-[[tris(hydroxymethyl)methyl]amino] ethane

15 sulphonic acid (TES) (0.9 mmoles), and gassed with 95% O_2 /5% CO_2 prior to use.

The isolated cells were first preincubated at 37°C for 30 minutes in a mixture containing 50 mM Tris-HCl buffer (pH 7.5), 10 mM potassium citrate, 10 mM $MgCl_2$, 1 mM dithiothreitol (DTT), and BSA (0.8 mg/ml). The reaction was then initiated by

20 the adding an aliquot of the preincubated cells to an assay mixture (final volume, 500 μ l) containing 50 mM Tris-HCl buffer (pH 7.5), 10 mM potassium citrate, 10 mM $MgCl_2$, 1 mM dithiothreitol (DTT), BSA (0.8 mg/ml), 3.75 mM ATP, 0.125 mM acetyl-CoA, and 12.5 mM $NaH[C^{14}]O_3$ (0.44 μ Ci/ μ mol). After incubation at 37°C for 10 minutes, the reaction was terminated with 0.1 ml of 6M HCl. The reaction mixture

25 was then allowed to stand in a vacuum desiccator for 30 minutes to remove the unreacted $NaH[C^{14}]O_3$ and followed by centrifugation at 1500 g for 10 minutes to eliminate the insoluble material. A 0.5 ml aliquot of the supernatant was taken and transferred to scintillation vials. The $[C^{14}]$ radioactivity was measured with a liquid scintillation counter. The remaining cell suspension was assayed for protein content.

30 Specific activity of the enzyme was expressed as mU/g cell dry weight, where 1 U

of acetyl-CoA carboxylase was defined as that amount which catalyzed the carboxylation of 1 μ mole acetyl-CoA per minute.

Assay for lipolytic activity

5 The lipolytic action of hGH 177-191 and analogues on the isolated adipose tissues was demonstrated by the release of glycerol and free fatty acid (FFA) into the medium during incubation at 37°C.

10 Adipose tissues were removed from animals and sliced into segments of approximately 200 mg each. Then the tissues were pre-incubated in 25 ml vials containing 2 ml of Krebs-Ringer bicarbonate (KRB) buffer, 4% defatted BSA and 5.5 mM glucose under an atmosphere of carbogen (95%O₂/ 5%CO₂) at 37°C for 1 hour. Tissues were then transferred to another vials with fresh medium and the incubation was initiated by adding hGH 177-191 peptide or analogues into the vials.

15 The mixtures were then incubated at 37°C for 90 minutes. After incubation, the tissues were removed and aliquots (200 μ l) of samples withdrawn from the medium were assayed for the contents of glycerol or free fatty acid (FFA) either by the enzyme assay (glycerol kinase) or colorimetric (copper-dye) spectrometry. The NADH or color produced was then monitored by the absorption at 340 nm and 610

20 nm, respectively.

Assay for Oxidation of Free Fatty Acid

 The effect of hGH 177-191 or analogues on the free fatty acid (FFA) oxidation in the adipose tissues is evaluated by the measure of converted [C¹⁴]O₂ from C¹⁴-

25 palmitic acid. The [C¹⁴]O₂ a final product of FFA oxidation, was trapped by hyamine hydroxide and measured by liquid scintillation counter. The rate of FFA oxidation was then determined by [C¹⁴] radioactivity.

 Adipose tissues removed from laboratory animals were sliced into segments

30 of approximately 200 mg each. The tissues in 25 ml vials containing 2 ml of Krebs-

Ringer phosphate (KRP) buffer, and 2% defatted bovine serum albumin (BSA) were pre-incubated at 37°C for 30 minutes under an atmosphere of carbogen (95%O₂/5%CO₂). Then the tissues were transferred to Konte flasks with fresh incubation medium with 0.15 mM sodium [C¹⁴]-palmitate (final [C¹⁴]-specific activity, 0.20 μCi/μmol) and hGH 177-191 peptide or analogues (1 - 1000 nM). A filter paper roll was placed in a well inside of flask and then the flask was seal with rubber septum stopper. The incubation at 37°C was lasted for 1 hour under an atmosphere of carbogen and was then terminated by injecting 250 μl of 4.5 M H₂SO₄ with a needle through the rubber septum into the medium of a flask and 250 μl of hyamine hydroxide was injected to the filter paper roll in the centre well. Flasks were incubated for a further 1 hour to complete the absorption of [C¹⁴]O₂ by hyamine hydroxide. The filter paper rolls were then removed and transferred to scintillation vials. The [C¹⁴] radioactivity was measured with a liquid scintillation counter. The rate of [C¹⁴]-palmitic acid oxidation to [C¹⁴]O₂ was calculated and expressed as μmol/g tissue/hr.

Assay for Lipogenic activity

The rate of the incorporation of exogenous [C¹⁴]-glucose into total lipid in adipose tissue was measured as the index of anti-lipogenic activity of hGH 177-191.

Adipose tissues were sliced into segments of approximately 200 mg each and then placed in Krebs-Ringer bicarbonate (KRB) buffer (pH 7.4) containing 2% defatted BSA, and glucose (0.1 mg/ml) and gassed with 95% O₂ - 5% CO₂ at 37 °C. After 1 hr preincubation, the tissues were transferred to another 2 ml of fresh media containing [C¹⁴]-glucose (final specific activity 0.05 μCi/μmol), and 0.3 μM of hGH 177-191 in the presence or absence of insulin (0.1 mU/ml) for a further 90 min incubation (conditions as above). Then the tissues were removed, washed thoroughly with KRB buffer and lipid was extracted with 5 ml of chloroform/methanol (2:1, v/v). The extraction solution was washed with 2 ml of MeOH-H₂O solution containing 0.1% MgCl₂. A 2.5 ml aliquot of the washed extraction solution was

taken and transferred to scintillation vials. The [C^{14}] radioactivity was measured with a liquid scintillation counter. The rates of total lipid synthesis were expressed as $\mu\text{mol } [C^{14}]\text{-glucose incorporated into lipid/g tissue/hr.}$

5 **Assays for diacylglycerol (DAG) release**

Diacylglycerol released from isolated adipose tissue or adipocytes was quantitated using a radioenzymic assay, employing E coli DAG kinase and defined mixed micelle conditions to solubilize DAG and allow its quantitative conversion to [^{33}p] phosphatidic acid in the presence of [^{33}p]- γ -ATP. Following a
10 number of extraction steps to remove unreacted [^{33}p]- γ -ATP, separation of [^{33}p] phosphatidic acid was achieved with the use of 1 ml Am-PrepTM minicolumns.

Statistical analysis.

The Student's t-test was used to analyse the results. All data are expressed
15 as the mean \pm SEM. *P* values of <0.05 are accepted as statistically significant.

RESULTS

The chronic treatment of the obese mice and rats with the synthetic hGH 177-
20 191 and analogues was evaluated by the measurements of a number of parameters including cumulative body weight gain and daily food consumption. During the treatment period, a clear reduction of cumulative body weight gain was observed in the hGH 177-191 treated male as well as female animals when compared with the appropriate control (**Figures 1A, 1B**). When the data were
25 analysed and expressed as daily body weight gain, the treated male animals reduced their body weight gain from 0.22 ± 0.03 to 0.16 ± 0.04 g/day and the female animals from 0.30 ± 0.02 to 0.22 ± 0.04 g/day. The average daily body weight gains of the both male and female treated animals showed approximately 27% lower than those of the appropriate control groups. However, no significant
30 difference in the average daily food consumption among the 4 groups was

observed (**Figures 2A, 2B**). These anti-obesity actions of various analogues, as represented by SEQ ID NO: 11 (**Figure 11A**) were observed in obese mice. The synthetic analogues control body weight gain without affecting the appetite of the treated animals (**Figure 11B**). The similar reduction of body weight gain was found also in Zucker fatty (fa/fa) rats during hGH 177-191 treatment *via* either daily intraperitoneal injection or intradermal implantation of a slow-release pellet (**Figures 3A, 3B**). The food consumption of the treated Zucker rats was unchanged throughout the treatment period (**Figures 4A, 4B**). These data clearly demonstrated that the chronic treatment with hGH 177-191 peptide reduced the body weight gain without affecting food consumption.

As indicated by the measurements of epididymal and parametrical fat pads, the treated mice significantly reduced their adipose tissue weights up to 20% in the males and 12% in the females as compared with the controls of the same sex (**Table 1**). Lipogenesis is subject to the supply of precursor metabolites such as glucose and acetate. The effect of hGH 177-191 or analogues was therefore determined by measuring the incorporation of [¹⁴C]-glucose into lipid in isolated adipose tissues. The peptide hGH 177-191 and analogues (**Table 5**) reduced lipogenic activity *in vitro* more than 25% as compared with the control in isolated tissues from fatty Zucker rats. Decrease in lipogenesis of the adipose tissue isolated from hGH 177-191 treated mice was evident (**Figures 5A, 5B**). The tissue lipogenic activity reduced from 2.80 ± 0.33 to 2.33 ± 0.21 pmol/mg tissue/min in male mice and from 3.36 ± 0.13 to 2.99 ± 0.21 pmol/mg tissue/min in female mice. The lipolytic activity in adipose tissues of the hGH 177-191 treated obese animals was found to increase significantly in both sexes (**Figures 6A, 6B**). These results are consistent with the reduction in adipose tissue mass and cumulative body weight gain previously observed.

Table 2 depicts the effect of the hGH 177-191 treatment on the profiles of circulating levels of triglyceride and cholesterol. The total cholesterol in plasma was

significantly reduced from 4.44 ± 0.56 to 3.52 ± 0.39 mmol/l in the male animals but the plasma levels of cholesterol in the treated female animals were only slightly lower than those of the control ones. On the other hand hGH 177-191 did not influence the plasma levels of triglyceride in both sexes. In the presence of hGH 177-191 and various analogues, the oxidation of fatty acids (**Figure 7**) and the release of glycerol (**Table 4**) in adipose tissues isolated from obese animals were enhanced. This is consistent with the increase in lipolytic activity of the hGH 177-191 treated adipose tissues. All these *in vivo* and *in vitro* actions on lipid metabolism by the synthetic hGH 177-191 and analogues appear to be the result of the stimulation of the release of the cellular messenger diacylglycerol (**Figure 8**) which in turn modulates the key lipolytic enzyme hormone-sensitive lipase (**Figure 9**) and lipogenic enzymes acetyl-CoA carboxylase (**Figures 10A, 10B**) in target organs.

TABLE 1 Effect of synthetic hGH 177-191 peptide on body weight and adipose tissue mass of obese mice after 18-day chronic treatment. Animals were given a daily i.p. injection of either hGH 177-191 (200 µg/kg body weight) or equivalent volume of saline as control. All data represent the mean \pm SEM of 6 animals (* $p < 0.1$; ** $p < 0.05$).

Item	Male		Female	
	Control	Treated	Control	Treated
Initial body wt.(g)	47.5 \pm 3.1	48.6 \pm 2.0	46.7 \pm 3.6	48.2 \pm 3.4
Final body wt.(g)	51.4 \pm 3.0	51.5 \pm 2.3	52.1 \pm 3.2	52.2 \pm 2.9
Body wt.gain(g) ^(a)	3.9 \pm 0.6	2.9 \pm 0.6*	5.4 \pm 0.4	4.0 \pm 0.6**
Adipose tissue (g) ^(b)	3.18 \pm 0.43	2.52 \pm 0.25**	3.67 \pm 0.54	3.26 \pm 0.25**

(a) The difference between the initial and final body weights were considered as body weight gain.

(b) The intact epididymal or parametrical fat pads were the representative adipose tissues.

TABLE 2 Effect of synthetic hGH 177-191 on plasma levels of triglyceride and total cholesterol in obese mice after 18-day chronic treatment. Blood samples were collected from the cut tips of the tails of the anaesthetised animals. Data represent the mean \pm SEM of 6 animals (* $p < 0.05$).

Item	Male		Female	
	Control	Treated	Control	Treated
Triglyceride (mmol/l)	0.63 \pm 0.26	0.58 \pm 0.16	0.41 \pm 0.19	0.38 \pm 0.11
Cholesterol (mmol/l)	4.44 \pm 0.56	3.52 \pm 0.39*	3.01 \pm 0.52	2.84 \pm 0.29

TABLE 3. Female C57BL/6J (ob/ob) mice aged 26 weeks were randomly divided into two groups (Sample number = 6 in each group). The mice were given a daily intraperitoneal (i.p.) injection of SEQ ID 11 (500 µg/kg body weight) or saline for 18 days. After 18 days, all animals were given saline for another 18 days.

Item	Treatment (Day 0-18)		Post-treatment (Day 20-36)	
Drug	SEQ ID: 11	Saline	SEQ ID: 11	Saline
Initial body weight (g)	54.74 ± 4.96	51.50 ± 2.61	54.70 ± 4.15	54.08 ± 2.77
Final body weight (g)	54.42 ± 4.66	53.72 ± 2.46	56.70 ± 4.45	56.14 ± 2.42
Body weight gain (g)	-0.32 ± 0.23	2.22 ± 0.22	2.00 ± 0.24	2.06 ± 0.25
Ave. food consumption (g/mouse/day)	4.82 ± 0.22	4.88 ± 0.29	5.03 ± 0.13	5.03 ± 0.14
Blood glucose (mM)	6.2 ± 0.8	5.8 ± 0.9	6.0 ± 1.0	5.7 ± 0.07
Triglyceride (mM)	0.49 ± 0.10	0.60 ± 0.11	0.54 ± 0.15	0.57 ± 0.08

TABLE 4. *In vitro* effect of peptide analogues on the glycerol release during the lipolysis. Adipose tissues were isolated from male Zucker fatty (fa/fa) rats (12-14 week-old) and incubated with different concentrations of peptide or saline. Each test group contains 6 samples.

Peptide concentration (µM)	Glycerol release (µmol/g tissue/hr)			
	Analogues			
	SEQ ID 1	SEQ ID 11	SEQ ID 16	SEQ ID 33
0	1.42 ± 0.04			
0.1	1.82 ± 0.02	1.80 ± 0.03	1.83 ± 0.11	1.75 ± 0.12
1.0	1.86 ± 0.12	1.88 ± 0.13	1.79 ± 0.07	1.94 ± 0.12

TABLE 5. *In vitro* effect of peptide analogues on the inhibition of lipogenesis. Isolated adipose tissues from male Zucker fatty (fa/fa) rats (12-14 week-old) were incubated with peptide (0.3 μ M) in KRB buffer containing exogenous insulin (0.1 mU/ml). The rate of [C^{14}]-glucose incorporation into [C^{14}]-lipid (nmol/g tissue/hr) was measured as the lipogenic activity of adipose tissues. Each test group contains 6 determinations.

Analogues SEQ ID No.	<i>In vitro</i> lipogenesis	
	nmol / g tissue / hr	Percentage of control
1	181 \pm 10	74.74
7	184 \pm 41	75.86
8	188 \pm 19	77.25
11	167 \pm 28	68.90
13	174 \pm 5	71.65
19	185 \pm 6	76.23
21	176 \pm 27	72.60
22	185 \pm 24	76.03
23	174 \pm 16	71.48
31	143 \pm 20	58.91
32	164 \pm 17	67.50
33	187 \pm 18	76.84
Buffer control	243 \pm 13	100.00

- 40 -

SEQUENCE LISTING

(1) GENERAL INFORMATION:

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JIANG, Woei-Jia

(ii) TITLE OF INVENTION: Treatment of Obesity

(iii) NUMBER OF SEQUENCES: 33

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(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:
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(A) APPLICATION NUMBER:USSN 08/340389
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- 41 -

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

Leu	Arg	Ile	Val	Gln	Cys	Arg	Ser	Val	Glu	Gly	Ser	Cys	Gly	Phe
1				5					10					15

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

AA1	Arg	AA2	Val	Gln	AA3	AA4	Ser	Val	Glu	AA5	AA6	AA3	AA5	AA7
1				5					10					15

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 17 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Xm	Leu	Arg	Ile	Val	Gln	Cys	Arg	Ser	Val	Glu	Gly	Ser	Cys	Gly	Phe	Xn
1				5					10						15	

- 42 -

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 17 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

X	Leu	Arg	Ile	Val	Gln	Cys	Arg	Ser	Val	Glu	Gly	Ser	Cys	Gly	Phe	Y
1				5					10					15		

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 13 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Leu	Arg	Ile	Val	Gln	Cys	A-Ser	Val	B-Gly	Ser	Cys	Gly	Phe
1				5					10			

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 13 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Leu	Arg	Ile	Val	Gln	Cys	B-Ser	Val	A-Gly	Ser	Cys	Gly	Phe
1				5					10			

- 43 -

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 16 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Lys	Leu	Arg	Ile	Val	Gln	Cys	Arg	Ser	Val	Glu	Gly	Ser	Cys	Gly	Phe
1				5					10					15	

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 17 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Lys	Lys	Leu	Arg	Ile	Val	Gln	Cys	Arg	Ser	Val	Glu	Gly	Ser	Cys	Gly	Phe
1				5					10					15		

- 44 -

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Ala	Arg	Ile	Val	Gln	Cys	Arg	Ser	Val	Glu	Gly	Ser	Cys	Gly	Phe
1				5					10				15	

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Leu	Arg	Ala	Val	Gln	Cys	Arg	Ser	Val	Glu	Gly	Ser	Cys	Gly	Phe
1				5					10				15	

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Leu	Arg	Lys	Val	Gln	Cys	Arg	Ser	Val	Glu	Gly	Ser	Cys	Gly	Phe
1				5					10				15	

- 45 -

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Leu	Arg	Ile	Val	Gln	D-Cys	Arg	Ser	Val	Glu	Gly	Ser	D-Cys	Gly	Phe
1				5					10				15	

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Leu	Arg	Ile	Val	Gln	Pen	Arg	Ser	Val	Glu	Gly	Ser	Pen	Gly	Phe
1				5					10				15	

(2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Leu	Arg	Ile	Val	Gln	D-Pen	Arg	Ser	Val	Glu	Gly	Ser	D-Pen	Gly	Phe
1				5					10				15	

- 46 -

(2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Leu	Arg	Ile	Val	Gln	Cys	D-Arg	Ser	Val	Glu	Gly	Ser	Cys	Gly	Phe
1				5					10				15	

(2) INFORMATION FOR SEQ ID NO:16:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Leu	Arg	Ile	Val	Gln	Cys	Lys	Ser	Val	Glu	Gly	Ser	Cys	Gly	Phe
1				5					10				15	

(2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Leu	Arg	Ile	Val	Gln	Cys	D-Lys	Ser	Val	Glu	Gly	Ser	Cys	Gly	Phe
1				5					10				15	

- 47 -

(2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

Leu	Arg	Ile	Val	Gln	Cys	Arg	Ser	Val	Glu	Ala	Ser	Cys	Ala	Phe
1				5					10				15	

(2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

Leu	Arg	Ile	Val	Gln	Cys	Arg	Ser	Val	Glu	D-Ala	Ser	Cys	D-Ala	Phe
1				5					10				15	

(2) INFORMATION FOR SEQ ID NO:20:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

Leu	Arg	Ile	Val	Gln	Cys	Arg	Ser	Val	Glu	Gly	Ala	Cys	Gly	Phe
1				5					10				15	

- 48 -

(2) INFORMATION FOR SEQ ID NO:21:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Leu	Arg	Ile	Val	Gln	Cys	Arg	Ser	Val	Glu	Gly	Ser	Cys	Ala	Phe
1				5					10					15

(2) INFORMATION FOR SEQ ID NO:22:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Leu	Arg	Ile	Val	Gln	Cys	Arg	Ser	Val	Glu	Gly	Ser	Cys	Gly	Ala
1				5					10					15

(2) INFORMATION FOR SEQ ID NO:23:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

Leu	Arg	Ile	Val	Gln	Cys	Lys	Ser	Val	Glu	Gly	Ser	Cys	Gly	Phe
1				5					10					15

- 49 -

(2) INFORMATION FOR SEQ ID NO:24:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

Leu	Arg	Ile	Val	Gln	Cys	Orn	Ser	Val	Glu	Gly	Ser	Cys	Gly	Phe
1				5					10					15

(2) INFORMATION FOR SEQ ID NO:25:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

Leu	Arg	Ile	Val	Gln	Cys	Lys	Ser	Val	Asp	Gly	Ser	Cys	Gly	Phe
1				5					10					15

(2) INFORMATION FOR SEQ ID NO:26:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

Leu	Arg	Ile	Val	Gln	Cys	Orn	Ser	Val	Asp	Gly	Ser	Cys	Gly	Phe
1				5					10					15

- 50 -

(2) INFORMATION FOR SEQ ID NO:27:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

Leu	Arg	Ile	Val	Gln	Cys	Glu	Ser	Val	Lys	Gly	Ser	Cys	Gly	Phe
1				5					10				15	

(2) INFORMATION FOR SEQ ID NO:28:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

Leu	Arg	Ile	Val	Gln	Cys	Glu	Ser	Val	Orn	Gly	Ser	Cys	Gly	Phe
1				5					10				15	

(2) INFORMATION FOR SEQ ID NO:29:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

Leu	Arg	Ile	Val	Gln	Cys	Asp	Ser	Val	Lys	Gly	Ser	Cys	Gly	Phe
1				5					10				15	

- 51 -

(2) INFORMATION FOR SEQ ID NO:30:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Leu Arg Ile Val Gln Cys Asp Ser Val Orn Gly Ser Cys Gly Phe
 1 5 10 15

(2) INFORMATION FOR SEQ ID NO:31:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 16 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

desaminoLeu Arg Ile Val Gln Cys Arg Ser Val Glu Gly Ser Cys Gly Phe
 1 5 10 15

(2) INFORMATION FOR SEQ ID NO:32:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 16 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

acetyl Leu Arg Ile Val Gln Cys Arg Ser Val Glu Gly Ser Cys Gly Phe
 1 5 10 15

- 52 -

(2) INFORMATION FOR SEQ ID NO:33:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 16 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

Leu	Arg	Ile	Val	Gln	Cys	Arg	Ser	Val	Glu	Gly	Ser	Cys	Gly	Phe	amide
1				5					10					15	

REFERENCES:

1. Ultsch, M.H., Somers, W., Kossiakof, A.A. and DeVos, A.M. (1994), *J. Mol. Biol.* **236**: 286-299.
2. Ng, F.M., Bornstein, J., Welker, C., Zimmet, P.Z. and Taft, P. (1974). *Diabetes* **23**: 943-949.
3. Frigeri, L.G., Teguh, C., Lind.N., Wolff, G.L. and Lewis, U.J. (1988). *Endocrinology* **122**: 2940-2945.
4. Moore, W.V., Moore, K.C., McLachlan, C.G., Fuller, N.J., Burnett, G.B. and Frane, J.W. (1988). *Endocrinology* **122**: 2920-2926.
5. Zeisel, H.J., Petrykowski, W.V. and Wais, U. (1992). *Horm. Res.* **37** (Suppl.2): 5-13.
6. Wabitsch, M. and Heinze, E. (1993). *Horm. Res.* **49**: 5-9.
7. Christman, G.M. and Halme, J.K. (1992) *Fertil. Steril.* **57**: 12-14.
8. Jacobs, H.S. (1992) *Horm. Res.* **38** (Suppl.1): 14-21.
9. Crist, D.M., Peake, G.T., Lofffield, R.B., Kraner, J.C. and Egan, P.A. (1991) *Mech. Ageing Dev.* **58**: 191-205.
10. Strobl, J.S. and Thomas, M.J. (1994) *Pharm. Rev.* **46**: 1-34.
11. Raben, M.S. and Hollenberg, C.H. (1959) *J. Clin. Invest.* **38**: 484-488.
12. Bengtsson, B.A., Eden, S., Lonn, L., Kvist, H., Stokland, A., Lindstedt, G., Bosaeus, I., Tolli, J., Sjostorm, L. and Isaksson, O.G.P. (1993) *J. Clin. Endocrinol. Metab.* **76**: 309-317.
13. Skaggs, S.R. and Crist, D.M. (1991) *Horm. Res.* **35**: 19-24.
14. Etherton, T.D., Wiggins, J.P., Evock, C.M., Chung, C.S., Rebhun, J.F., Walton, P.E. and Steele, N.C. (1987). *J. Anim. Sci.* **64**: 433-443.
15. Jiang, W.J., Shih, I.L., Tsai, H., Huang, Y.T. and Koh, T.J. (1993). 13th American Peptide Symposium, Edmonton, Canada, P-334 (Abstract).
16. Gertner, J.M. (1993) *Horm. Res.* **40**: 10-15.
17. Davidson, M.B. (1987) *Endocrine Rev.* **8**: 115-131.

18. Tai, P.K., Liao, J.F., Chen, E.H., Dietz, J., Schwards, J. and Carter-Su, C. (1990) *J. Biol. Chem.* **265**: 21828-21834.
19. Dobo, M. (1975) PhD. Thesis, Department of Biochemistry, Monash University, Australia.
20. McNeillie, E.M. and Zammit, V.A. (1982) *Biochem. J.* **204**: 273-288.
21. Gertner, J.M. (1992) *Horm. Res.* **38** (Suppl.2): 41-43.
22. Ng, F.M., Adamafio, N.A. and Graystone, J.E. (1990) *J. Mol. Endocrinol.* **4**: 43-49.
23. Ng, F.M. and Heng, D. (1988) *Asia Pacific Commun. Biochem.* **2**: 47-51.
24. Fossati, P., Prencipe, L. (1982) *Clin. Chem.* **28**: 2077-80.
25. Allain, C.C., Poon, L.S., Chan, C.S.G., Richmond, W. and Fu, P.C. (1974) *Clin. Chem.* **20**: 470-475.

VARIOUS ASPECTS OF THE INVENTION ARE SUMMARISED AS FOLLOWS:

1. A method for the treatment of obesity in an animal, which comprises administering to the animal an effective amount of a peptide which comprises the carboxyl-terminal sequence of a growth hormone, wherein said peptide is not the intact full-length growth hormone.
2. A method according to claim 1, wherein the animal is a human.
3. A method according to claim 1 or claim 2, wherein the peptide comprises the carboxyl-terminal sequence of human growth hormone.
4. A method according to claim 3, wherein the peptide comprises the carboxyl-terminal sequence of human growth hormone containing amino acid residues 177-191,
Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
177 178 179 180 181 182 183 184 185 186 187 188 189 190 191
(SEQ ID NO: 1)
a cyclic disulfide thereof or an organic or inorganic acid addition salt thereof.
5. A method according to claim 1 or claim 2, wherein the peptide comprises a homologue, analogue, mutant, variant or derivative obtained by elongation, insertion, deletion or substitution of amino acids in, or introduction of a cyclic amide bond between the side chains of amino acids of, chemical modification of, the native carboxyl-terminal sequence of human growth hormone containing amino acid residues 177-191, a cyclic disulfide thereof or an organic or inorganic acid addition salt thereof.

6. A method according to claim 5, wherein the peptide comprises an analogue of the sequence:

AA1-Arg-AA2-Val-Gln-AA3-AA4-Ser-Val-Glu-AA5-AA6-AA3-AA5-AA7
(SEQ ID NO: 2)

wherein:

AA1 is selected from the group consisting of L- or D- Leu and Ala
AA2 is selected from the group consisting of L- or D- Ile, Lys and Ala
AA3 is selected from the group consisting of L- or D- Cys and Pen
AA4 is selected from the group consisting of L- or D- Arg and Lys
AA5 is selected from the group consisting of L- or D- Ala and Gly
AA6 is selected from the group consisting of L- or D- Ser and Ala
AA7 is selected from the group consisting of L- or D- Phe and Ala

or a cyclic disulfide thereof or an organic or inorganic acid addition salt thereof.

7. A method according to claim 5, wherein the peptide comprises an analogue of the sequence:

X_m-Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe-X_n
(SEQ ID NO: 3)

wherein X is selected from the group consisting of L- or D- Arg, His, Lys and Tyr and m and n are each 0, 1, 2 or 3 with the provision that at least m or n is 1,

a cyclic disulfide thereof or an organic and inorganic acid addition salt thereof.

8. A method according to claim 5, wherein the peptide comprises an analogue of the sequence:

X-Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe-Y
(SEQ ID NO: 4)

wherein X is selected from the group consisting of the desamino form (H), CH₃CO- and other acyl groups; and Y is selected from the group of -COOH, -CONH₂ and alkyl amide groups,

a cyclic disulfide thereof or an organic or inorganic acid addition salt thereof.

9. A method according to claim 5, wherein the peptide comprises an analogue having a cyclic amide bond between amino acids A and B of the sequence:

Leu-Arg-Ile-Val-Gln-Cys-A-Ser-Val-B-Gly-Ser-Cys-Gly-Phe (SEQ ID NO:5)

Leu-Arg-Ile-Val-Gln-Cys-B-Ser-Val-A-Gly-Ser-Cys-Gly-Phe (SEQ ID NO:6)

wherein A is selected from the group consisting of L- or D- Lys and Orn; and B is selected from the group consisting of L- or D- Glu and Asp,

a cyclic disulfide thereof or an organic or inorganic acid addition salt thereof.

10. A method according to claim 5, wherein the peptide comprises an analogue selected from:

- 58 -

- (i) Lys-Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 7)
- (ii) Lys-Lys-Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 8)
- (iii) Ala-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 9)
- (iv) Leu-Arg-Ala-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 10)
- (v) Leu-Arg-Lys-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 11)
- (vi) Leu-Arg-Ile-Val-Gln-D-Cys-Arg-Ser-Val-Glu-Gly-Ser-D-Cys-Gly-Phe
(SEQ ID NO: 12)
- (vii) Leu-Arg-Ile-Val-Gln-Pen-Arg-Ser-Val-Glu-Gly-Ser-Pen-Gly-Phe
(SEQ ID NO: 13)
- (viii) Leu-Arg-Ile-Val-Gln-D-Pen-Arg-Ser-Val-Glu-Gly-Ser-D-Pen-Gly-Phe
(SEQ ID NO: 14)
- (ix) Leu-Arg-Ile-Val-Gln-Cys-D-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 15)
- (x) Leu-Arg-Ile-Val-Gln-Cys-Lys-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 16)
- (xi) Leu-Arg-Ile-Val-Gln-Cys-D-Lys-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 17)
- (xii) Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Ala-Ser-Cys-Ala-Phe
(SEQ ID NO: 18)
- (xiii) Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-D-Ala-Ser-Cys-D-Ala-Phe
(SEQ ID NO: 19)
- (xiv) Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ala-Cys-Gly-Phe
(SEQ ID NO: 20)
- (xv) Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Ala-Phe
(SEQ ID NO: 21)
- (xvi) Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Ala
(SEQ ID NO: 22)
- (xvii) Leu-Arg-Ile-Val-Gln-Cys-Lys-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 23)
- (xviii) Leu-Arg-Ile-Val-Gln-Cys-Orn-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 24)
- (xix) Leu-Arg-Ile-Val-Gln-Cys-Lys-Ser-Val-Asp-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 25)
- (xx) Leu-Arg-Ile-Val-Gln-Cys-Orn-Ser-Val-Asp-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 26)
- (xxi) Leu-Arg-Ile-Val-Gln-Cys-Glu-Ser-Val-Lys-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 27)
- (xxii) Leu-Arg-Ile-Val-Gln-Cys-Glu-Ser-Val-Orn-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 28)
- (xxiii) Leu-Arg-Ile-Val-Gln-Cys-Asp-Ser-Val-Lys-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 29)
- (xxiv) Leu-Arg-Ile-Val-Gln-Cys-Asp-Ser-Val-Orn-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 30)
- (xxv) desaminoLeu-hGH 177-191 (SEQ ID NO: 31)
- (xxvi) acetyl-hGH 177-191 (SEQ ID NO: 32)
- (xxvii) hG H 177-191-amide (SEQ ID NO: 33)

wherein the amino acid residue abbreviations used are in accordance with the standard peptide nomenclature; a cyclic disulfide thereof or an organic or inorganic acid addition salt thereof; or in the case of analogues SEQ ID NO: 23 to SEQ ID NO: 30 an analogue wherein there is a cyclic amide bond between amino acid residues in bold.

11. A pharmaceutical composition for use in the treatment of obesity in an animal, comprising an effective amount of a peptide which comprises the carboxyl-terminal sequence of a growth hormone together with one or more pharmaceutically acceptable carriers and/or diluents, wherein said peptide is not the intact, full-length growth hormone.
12. A composition according to claim 11, wherein the peptide comprises the carboxyl-terminal sequence of human growth hormone.
13. A composition according to claim 12, wherein the peptide comprises the carboxyl-terminal sequence of human growth hormone containing amino acid residues 177-191,
Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SAQ ID NO: 1)
a cyclic disulfide thereof or an organic or inorganic acid addition salt thereof.
14. A composition according to claim 11, wherein the peptide comprises a homologue, analogue, mutant, variant or derivative obtained by elongation, insertion, deletion or substitution of amino acids in, chemical modification of, or introduction of a cyclic amide bond between the side chains of amino acids of the native carboxyl-terminal sequence of human growth hormone containing amino acid residues 177-191, a

cyclic disulfide thereof or an organic or inorganic acid addition salt thereof.

15. A composition according to claim 14, wherein the peptide comprises an analogue of the sequence:

AA1-Arg-AA2-Val-Gln-AA3-AA4-Ser-Val-Glu-AA5-AA6-AA3-AA5-AA7

(SEQ ID NO: 2)

wherein:

AA1 is selected from the group consisting of L- or D- Leu and Ala

AA2 is selected from the group consisting of L- or D- Ile, Lys and Ala

AA3 is selected from the group consisting of L- or D- Cys and Pen

AA4 is selected from the group consisting of L- or D- Arg and Ala

AA5 is selected from the group consisting of L- or D- Ala and Gly

AA6 is selected from the group consisting of L- or D- Ser and Ala

AA7 is selected from the group consisting of L- or D- Phe and Ala

or a cyclic disulfide thereof or an organic or inorganic acid addition salt thereof.

16. A composition according to claim 13, wherein the peptide comprises an analogues of the sequence:

Xm-Leu-Arg-Ile-Val-Gln-Cys-Arg -Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe-Xn

(SEQ ID NO: 3)

wherein X is selected from the group consisting of L- or D- Arg, His, Lys and Tyr, and m and n are each 0, 1, 2 or 3 with the provision that at least

m or n is 1, a cyclic disulfide thereof or an organic or inorganic and addition salt thereof.

17. A composition according to claim 14, wherein the peptide comprises an analogue of the sequence:

X-Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe-Y

(SEQ ID NO: 4)

wherein X is selected from the group consisting of the desamino form (H), CH₃CO- and other acyl groups; and Y is selected from the group of -COOH, -CONH₂ and alkyl amide groups,

a cyclic disulfide thereof or an organic or inorganic acid addition salt thereof.

18. A composition according to claim 14, wherein the peptide comprises an analogue having a cyclic amide bond between amino acids A and B of the sequence:

Leu-Arg-Ile-Val-Gln-Cys-A-Ser-Val-B-Gly-Ser-Cys-Gly-Phe

(SEQ ID NO: 5)

Leu-Arg-Ile-Val-Gln-Cys-B-Ser-Val-A-Gly-Ser-Cys-Gly-Phe

(SEQ ID NO: 6)

wherein A is selected from the group consisting of L- or D- Lys and Orn; and B is selected from the group consisting of L- or D- Glu and Asp,

a cyclic disulfide thereof or an organic or inorganic acid addition salt thereof.

19. A composition according to claim 13, wherein the peptide comprises an analogue selected from:

- (i) Lys-Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 7)
- (ii) Lys-Lys-Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 8)
- (iii) Ala-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 9)
- (iv) Leu-Arg-Ala-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 10)
- (v) Leu-Arg-Lys-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 11)
- (vi) Leu-Arg-Ile-Val-Gln-D-Cys-Arg-Ser-Val-Glu-Gly-Ser-D-Cys-Gly-Phe
(SEQ ID NO: 12)
- (vii) Leu-Arg-Ile-Val-Gln-Pen-Arg-Ser-Val-Glu-Gly-Ser-Pen-Gly-Phe
(SEQ ID NO: 13)
- (viii) Leu-Arg-Ile-Val-Gln-D-Pen-Arg-Ser-Val-Glu-Gly-Ser-D-Pen-Gly-Phe
(SEQ ID NO: 14)
- (ix) Leu-Arg-Ile-Val-Gln-Cys-D-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 15)
- (x) Leu-Arg-Ile-Val-Gln-Cys-Lys-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 16)
- (xi) Leu-Arg-Ile-Val-Gln-Cys-D-Lys-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 17)
- (xii) Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Ala-Ser-Cys-Ala-Phe
(SEQ ID NO: 18)
- (xiii) Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-D-Ala-Ser-Cys-D-Ala-Phe
(SEQ ID NO: 19)
- (xiv) Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ala-Cys-Gly-Phe
(SEQ ID NO: 20)
- (xv) Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Ala-Phe
(SEQ ID NO: 21)
- (xvi) Leu-Arg-Ile-Val-Gln-Cys-Arg-Ser-Val-Glu-Gly-Ser-Cys-Gly-Ala
(SEQ ID NO: 22)
- (xvii) Leu-Arg-Ile-Val-Gln-Cys-Lys-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 23)
- (xviii) Leu-Arg-Ile-Val-Gln-Cys-Orn-Ser-Val-Glu-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 24)
- (xix) Leu-Arg-Ile-Val-Gln-Cys-Lys-Ser-Val-Asp-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 25)
- (xx) Leu-Arg-Ile-Val-Gln-Cys-Orn-Ser-Val-Asp-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 26)
- (xxi) Leu-Arg-Ile-Val-Gln-Cys-Glu-Ser-Val-Lys-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 27)
- (xxii) Leu-Arg-Ile-Val-Gln-Cys-Glu-Ser-Val-Orn-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 28)
- (xxiii) Leu-Arg-Ile-Val-Gln-Cys-Asp-Ser-Val-Lys-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 29)
- (xxiv) Leu-Arg-Ile-Val-Gln-Cys-Asp-Ser-Val-Orn-Gly-Ser-Cys-Gly-Phe
(SEQ ID NO: 30)

(xxv)	desaminoLeu-hGH 177-191	(SEQ ID NO: 31)
(xxvi)	acetyl-hGH 177-191	(SEQ ID NO: 32)
(xxvii)	hGH 177-191-amide	(SEQ ID NO: 33)

wherein the amino acid residue abbreviations used are in accordance with the standard peptide nomenclature; a cyclic disulfide thereof or an organic or inorganic acid addition salt thereof; or in the case of analogues SEQ ID NO: 23 to SEQ ID NO: 30 an analogue wherein there is a cyclic amide bond between amino acid residues in bold.

Dated this 8th day of September 1997

Monash University metabolic pharmaceuticals LTD

By its Patent Attorneys

Davies Collison Cave



Effect of hGH 177-191 on the body weight gain of male C57BL/6J (ob/ob) mice by ip injection

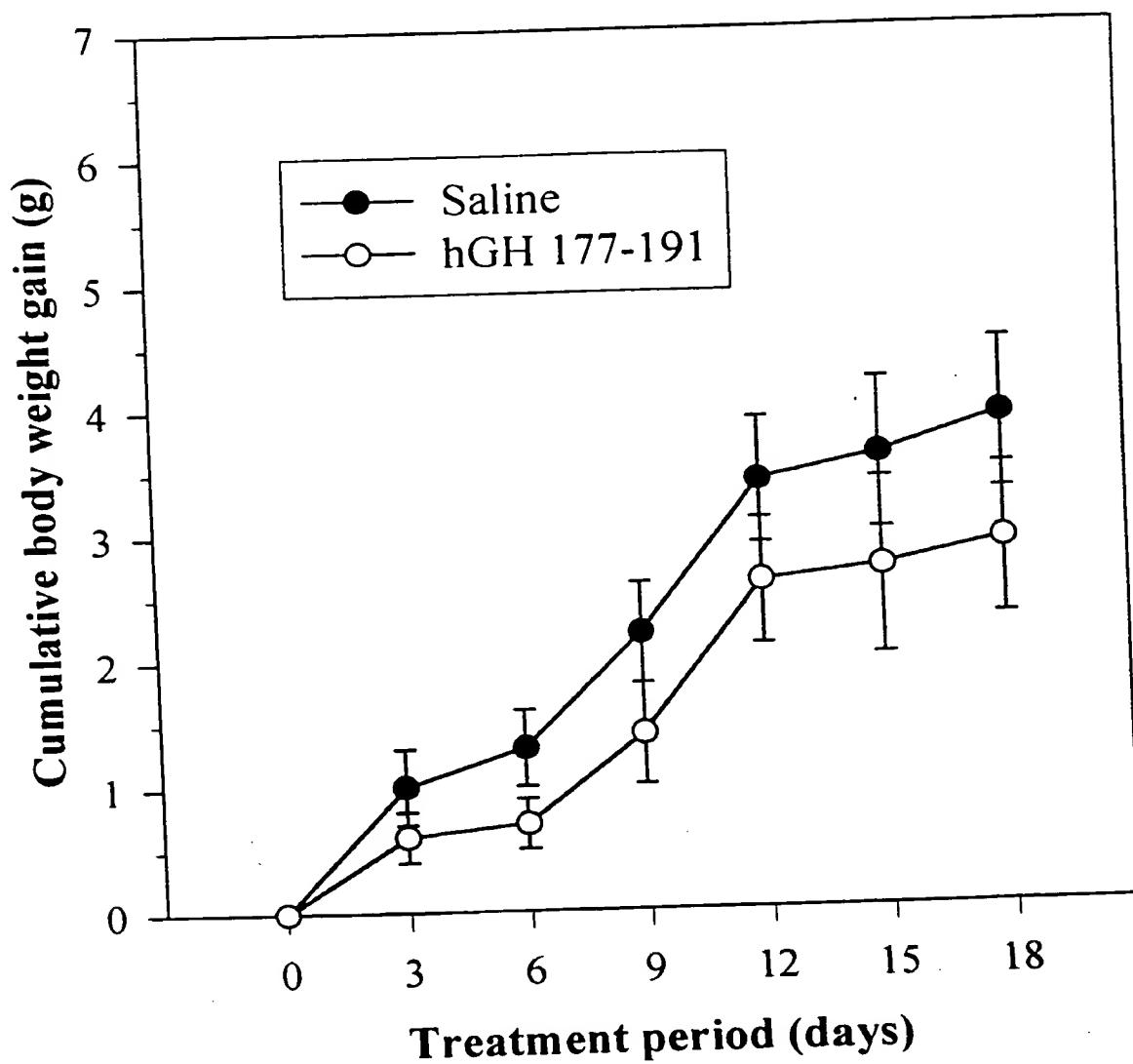


Figure 1A

Effect of hGH 177-191 on the body weight gain of female C57BL/6J (ob/ob) mice by ip injection

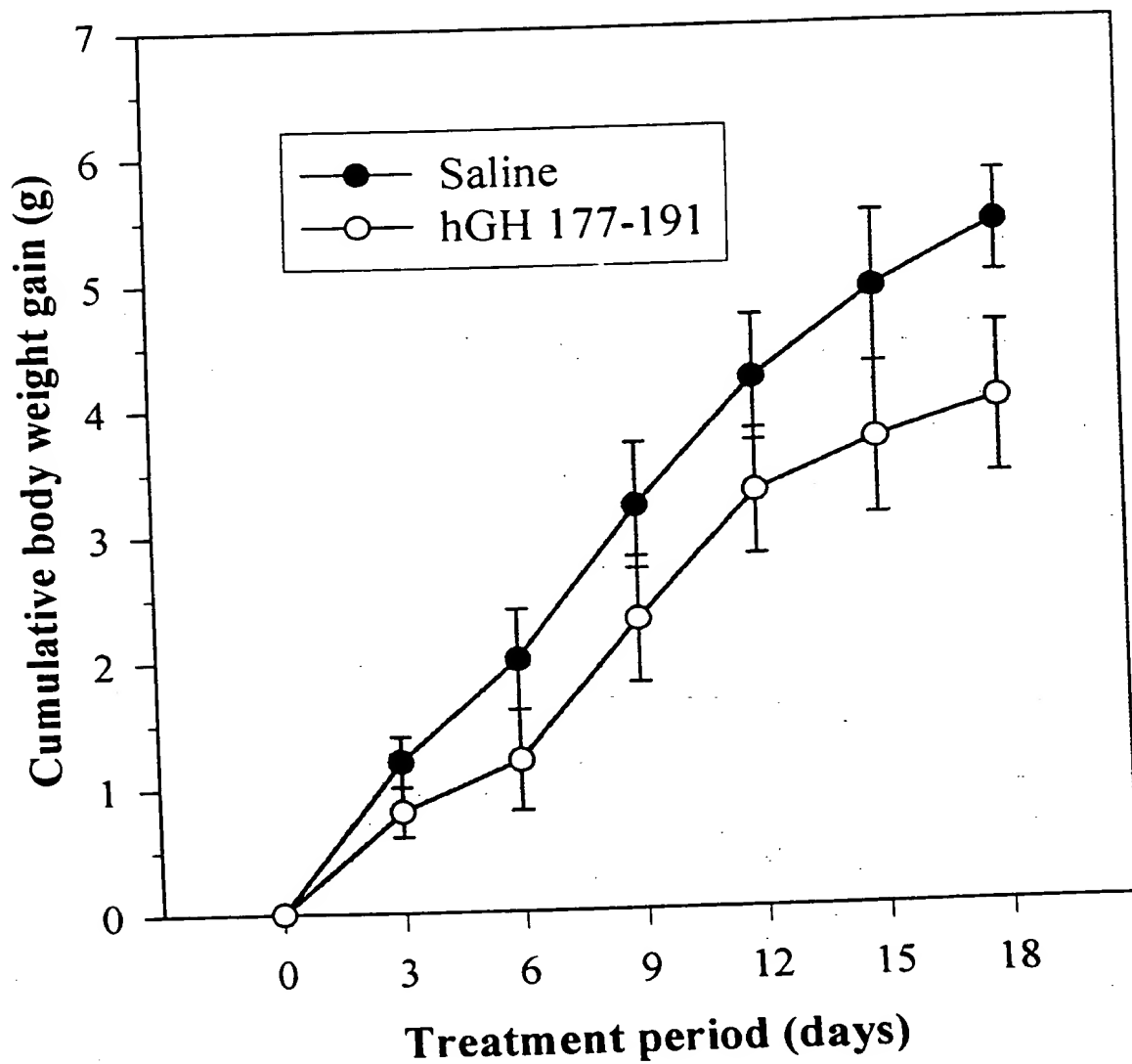


Figure 1B

Average daily food consumption of male C57BL/6J (ob/ob) mice during ip injection with hGH 177-191

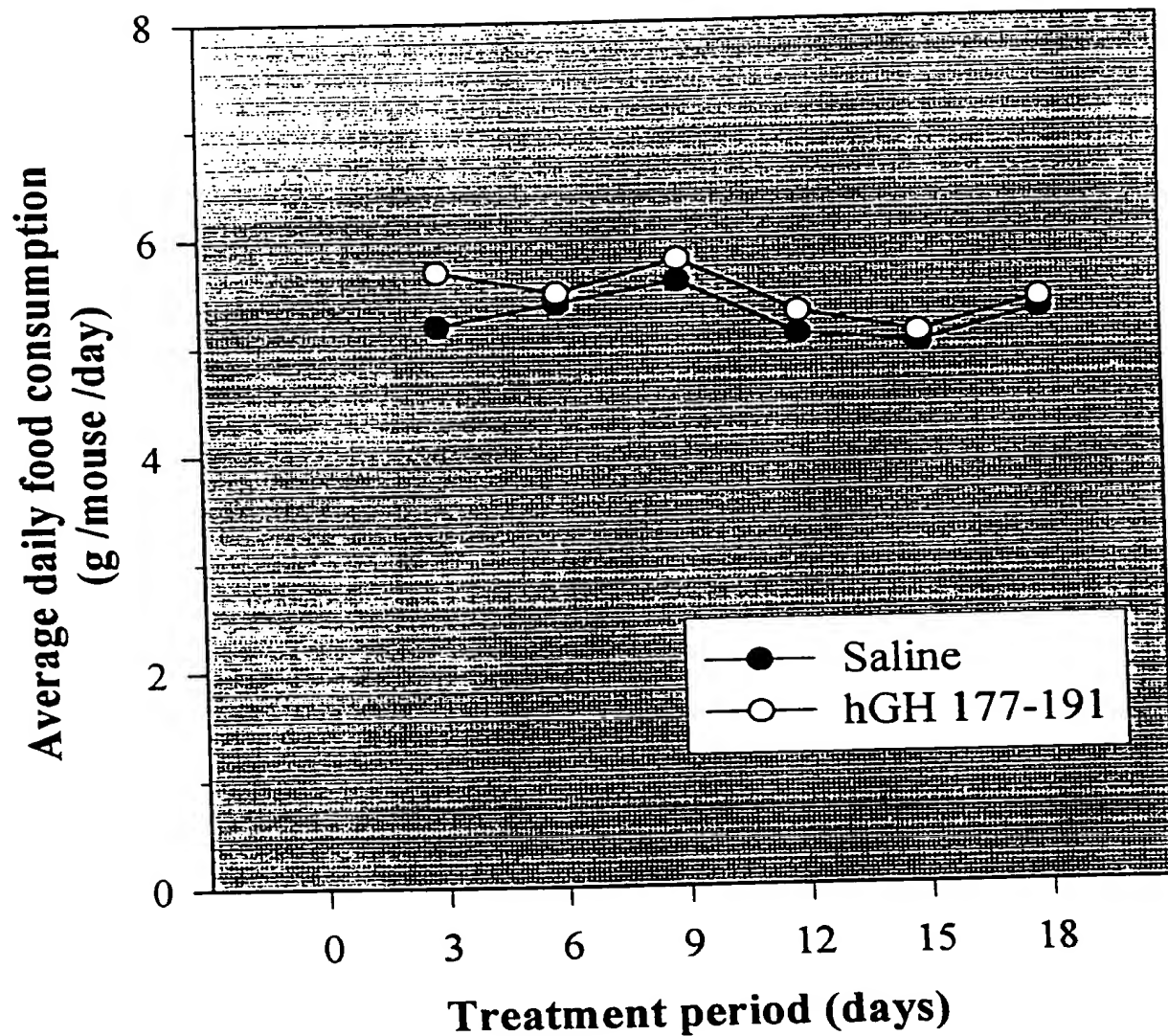


Figure 2A

Average daily food consumption of female C57BL/6J (ob/ob) mice during ip injection with hGH 177-191

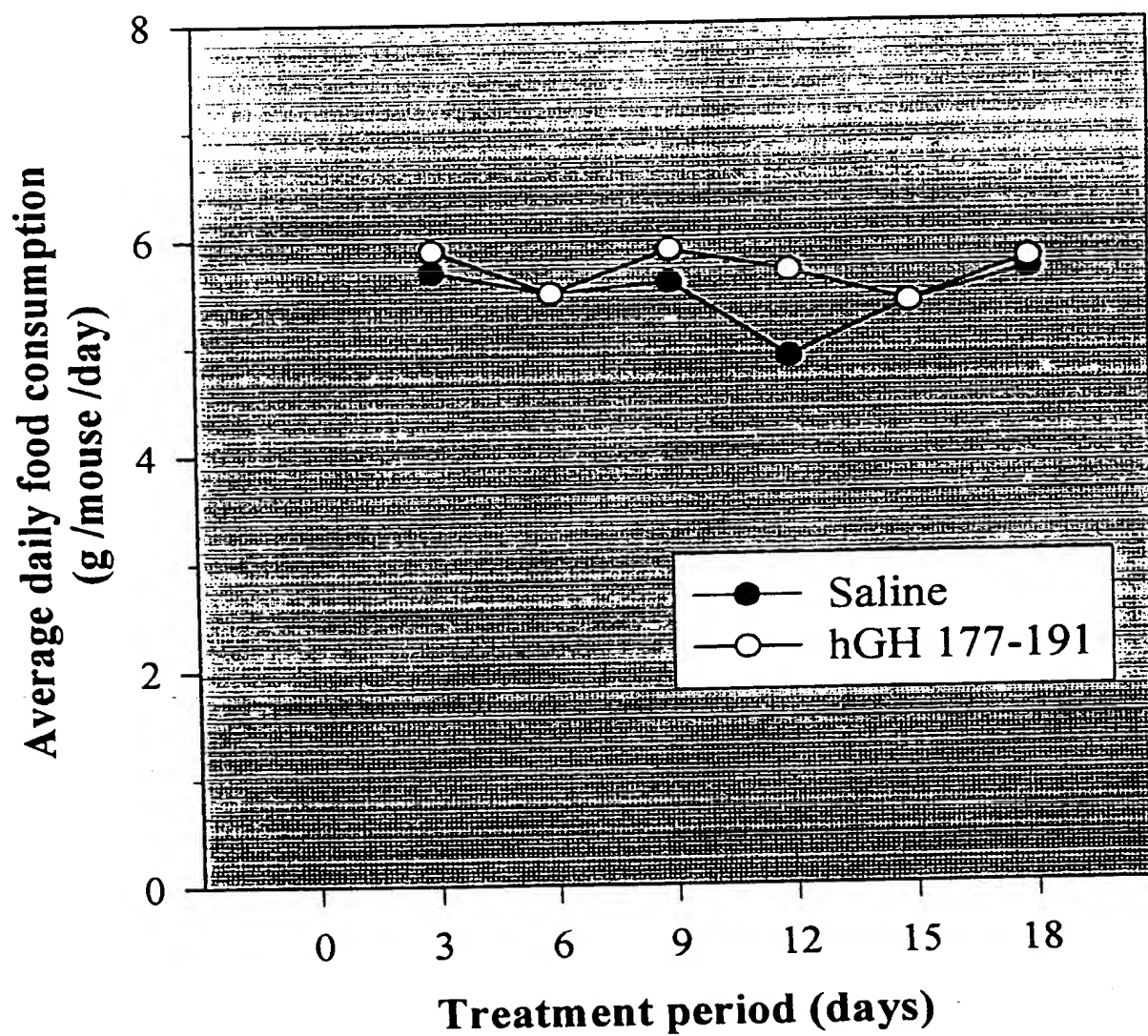


Figure 2B

**Effect of hGH 177-191 on the body weight gain
of Zucker Fatty (fa/fa) rats by ip injection**

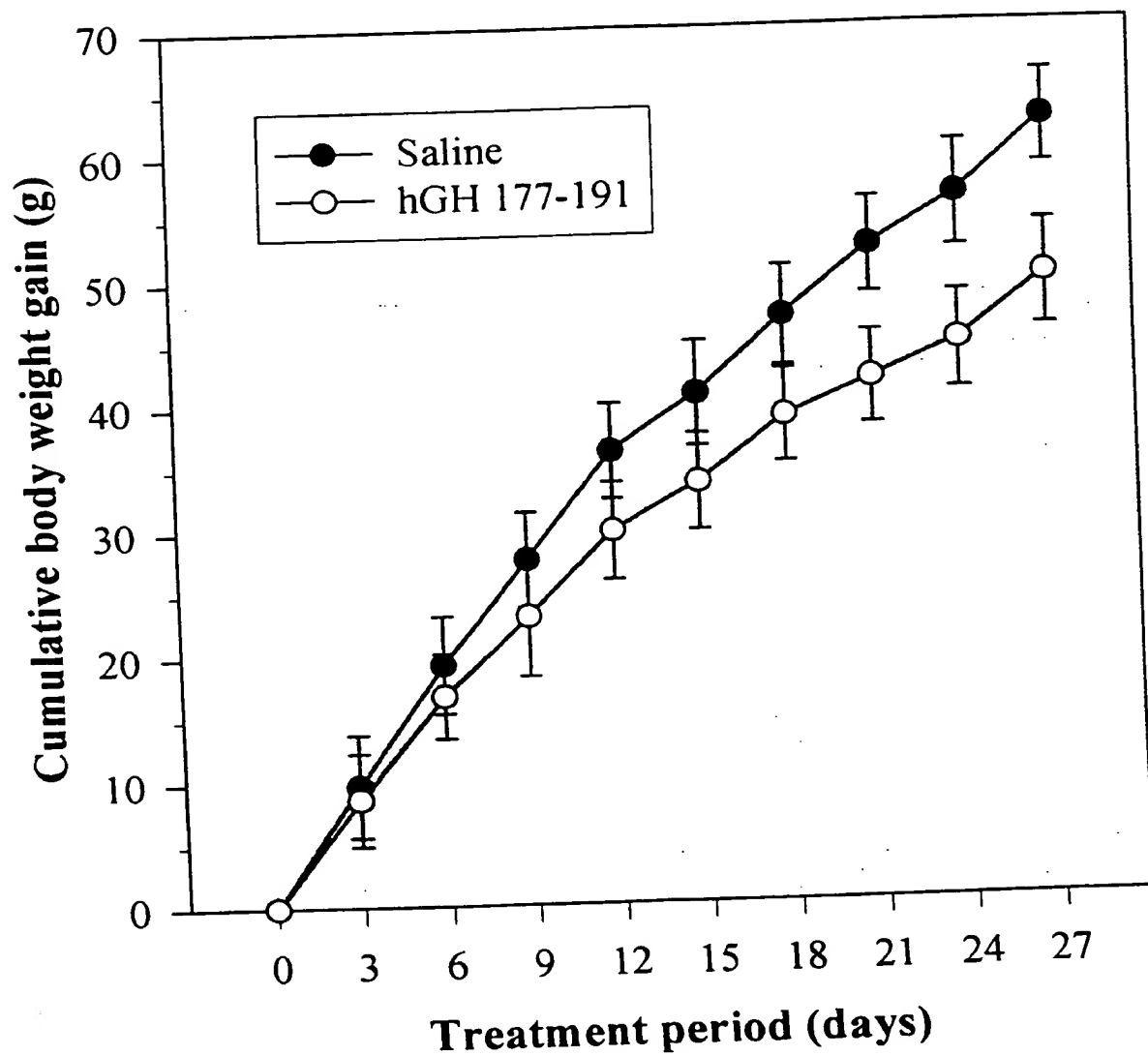


Figure 3A

**Effect of hGH 177-191 on the body weight gain
of Zucker Fatty (fa/fa) rats by id implant**

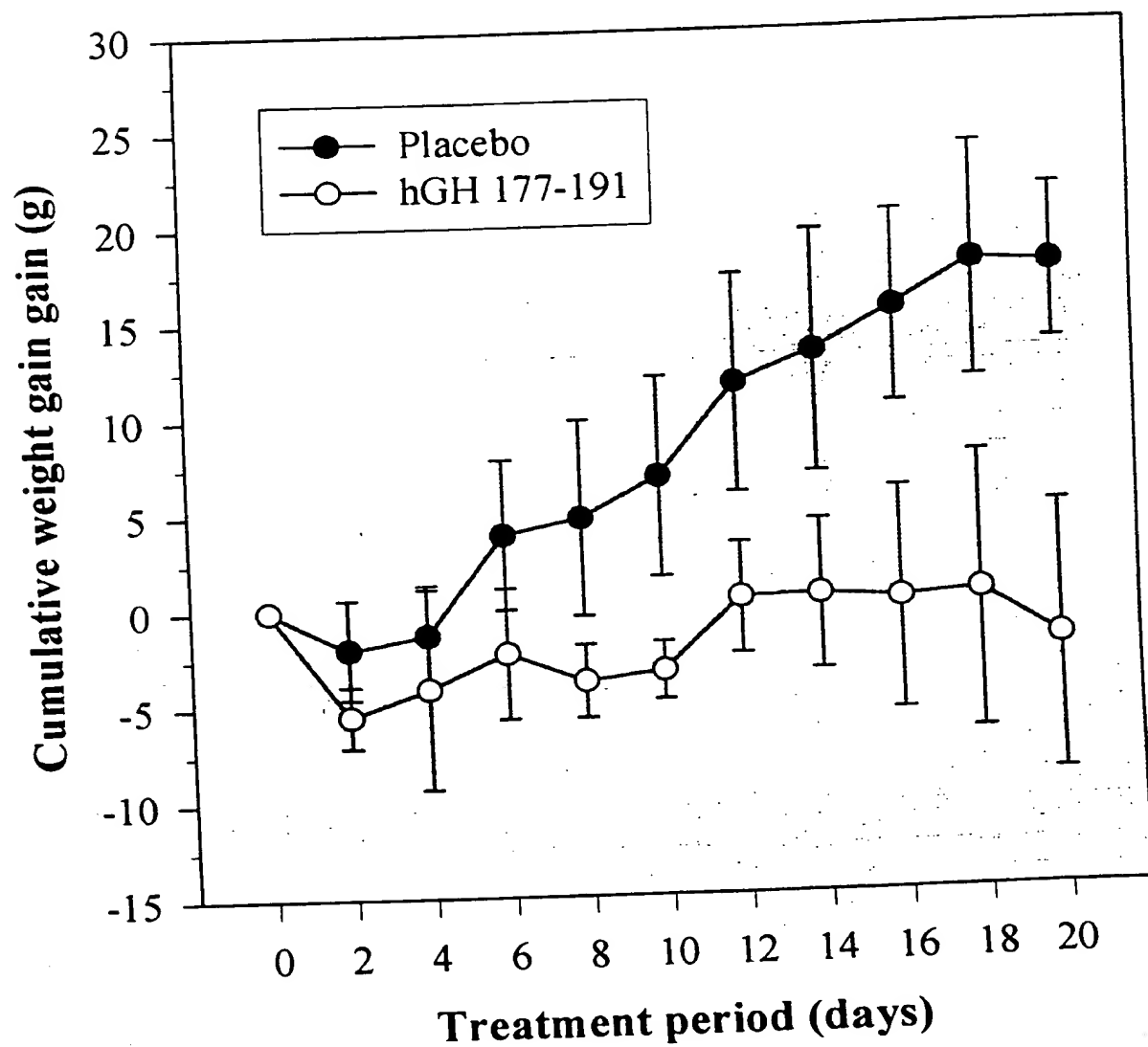


Figure 3B

Average daily food consumption of Zucker Fatty (fa/fa) rats during ip injection with hGH 177-191

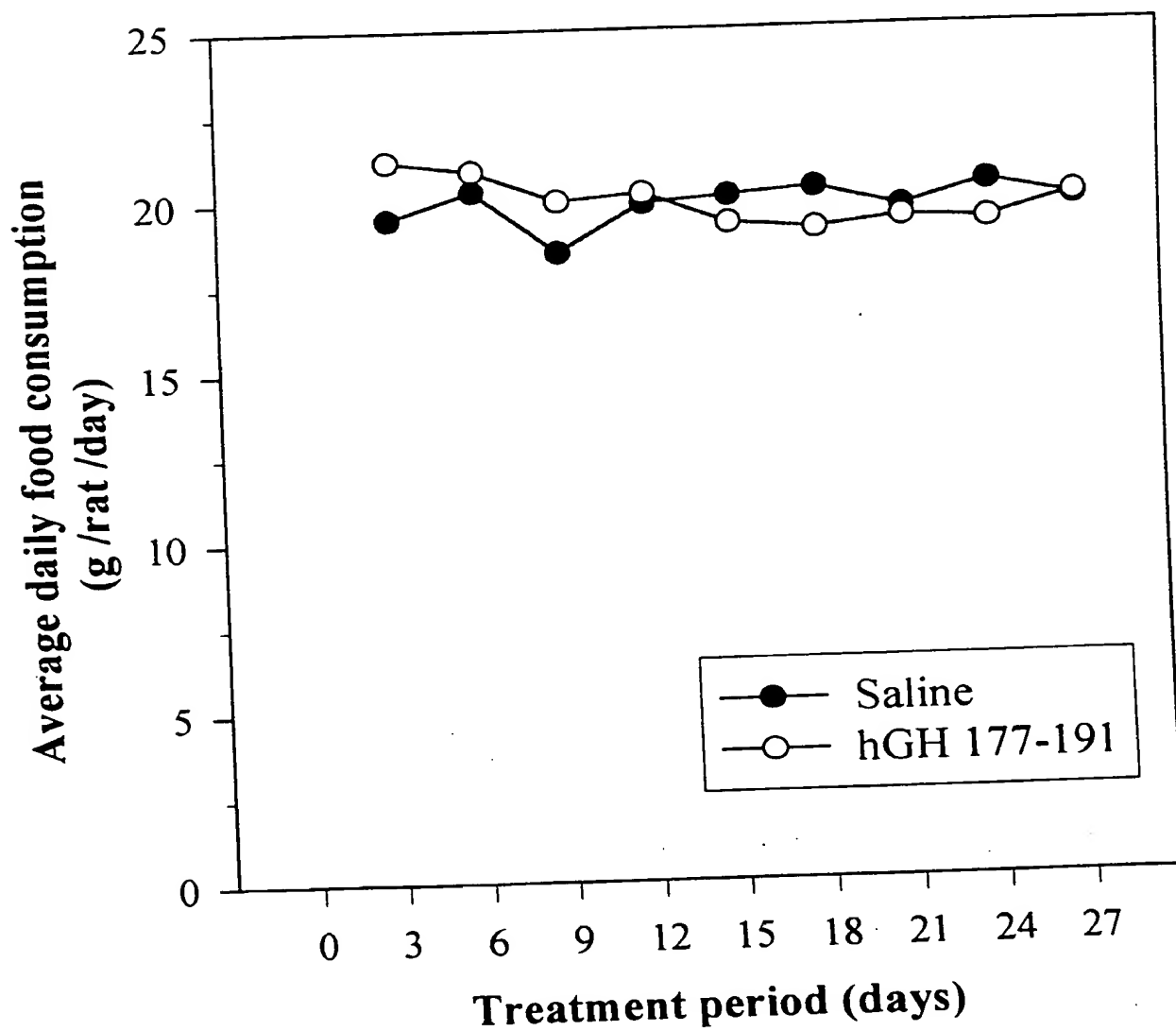


Figure 4A

Average daily food consumption of Zucker Fatty (fa/fa) rats during id implant with hGH 177-191

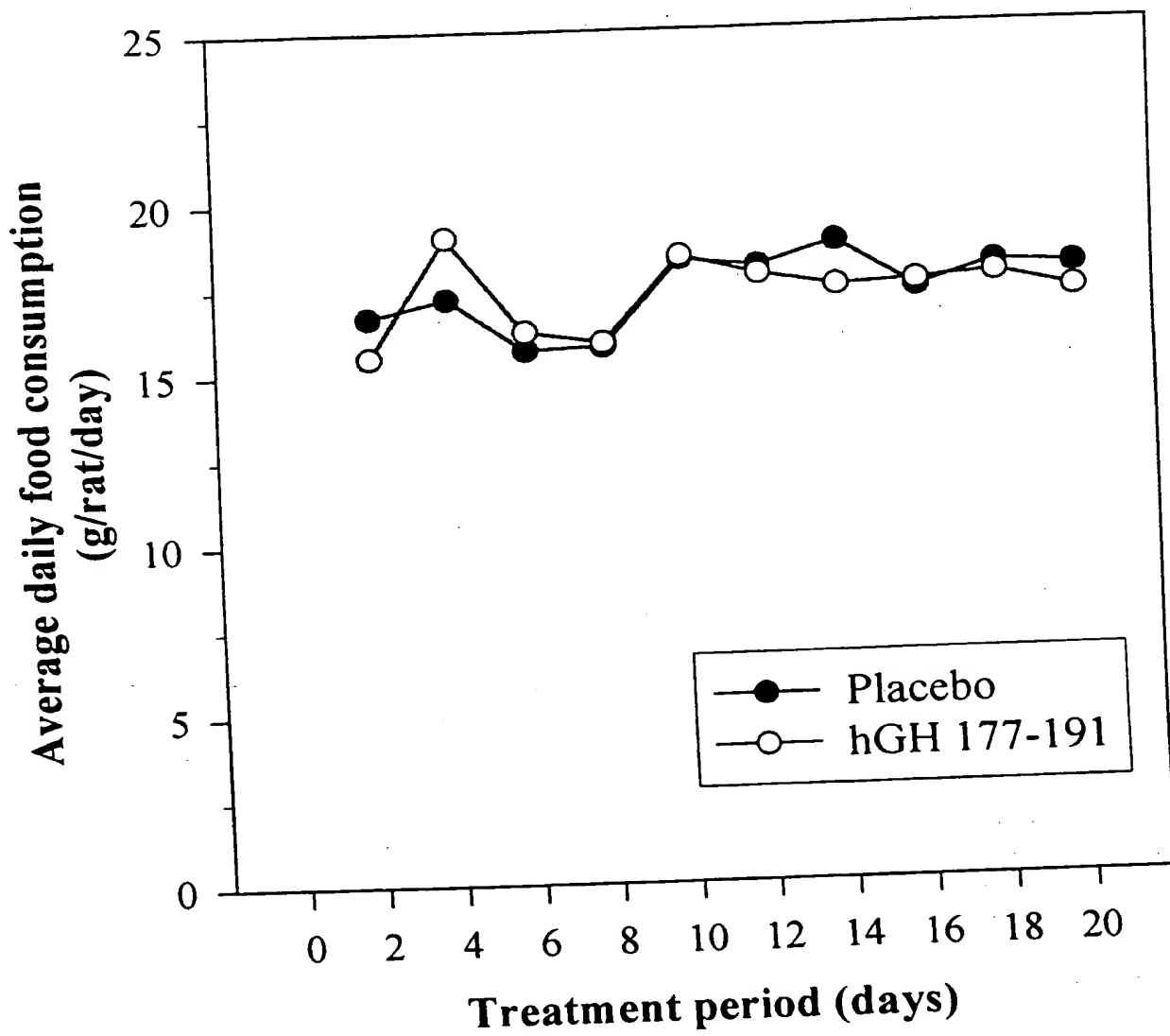


Figure 4B

***Ex vivo* effect on the lipogenesis in adipose tissues of the male C57BL/6J (ob/ob) mice after 18-day treatment**

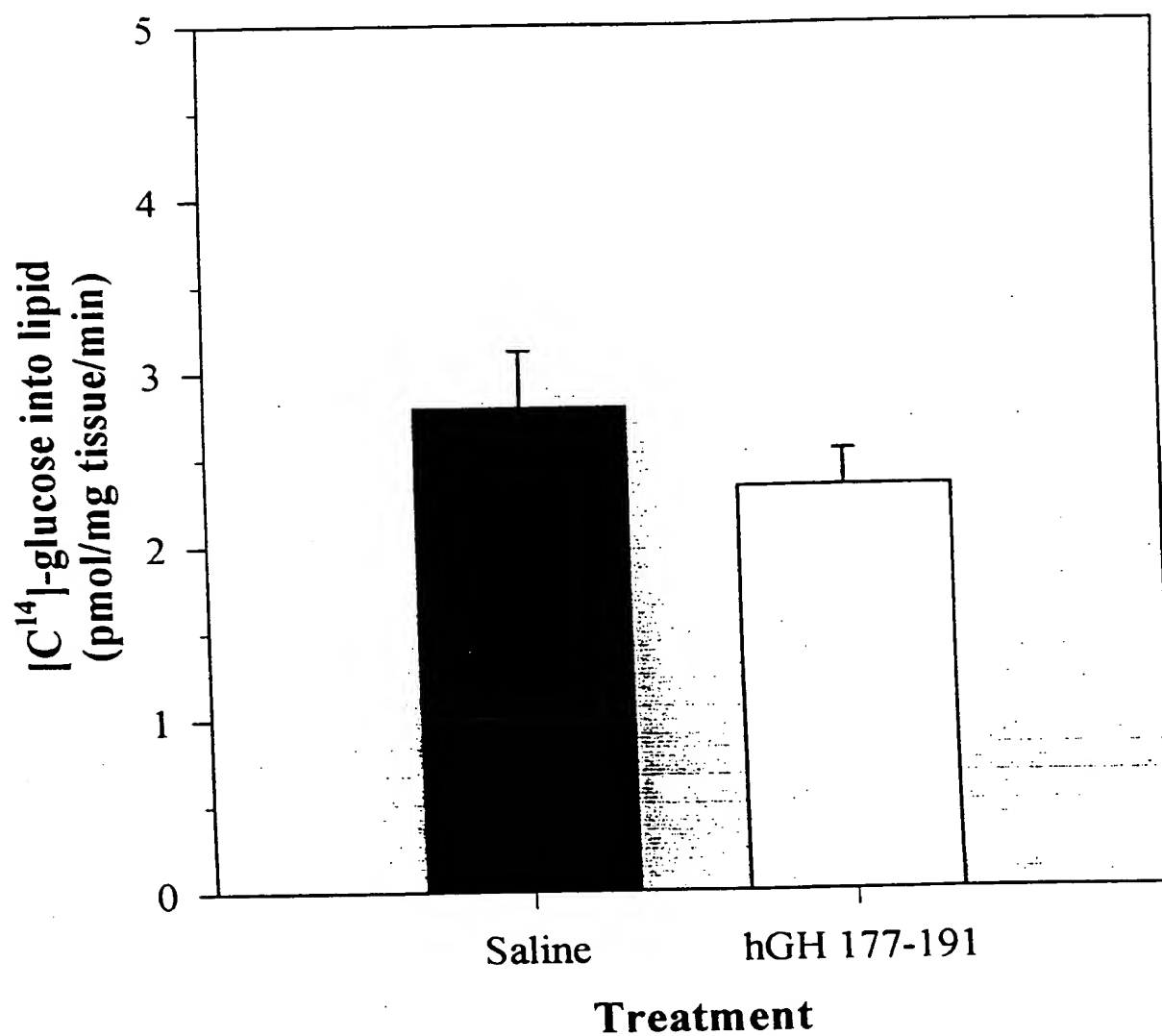


Figure 5A

***Ex vivo* effect on the lipogenesis in adipose tissues of the female C57BL/6J (ob/ob) mice after 18-day treatment**

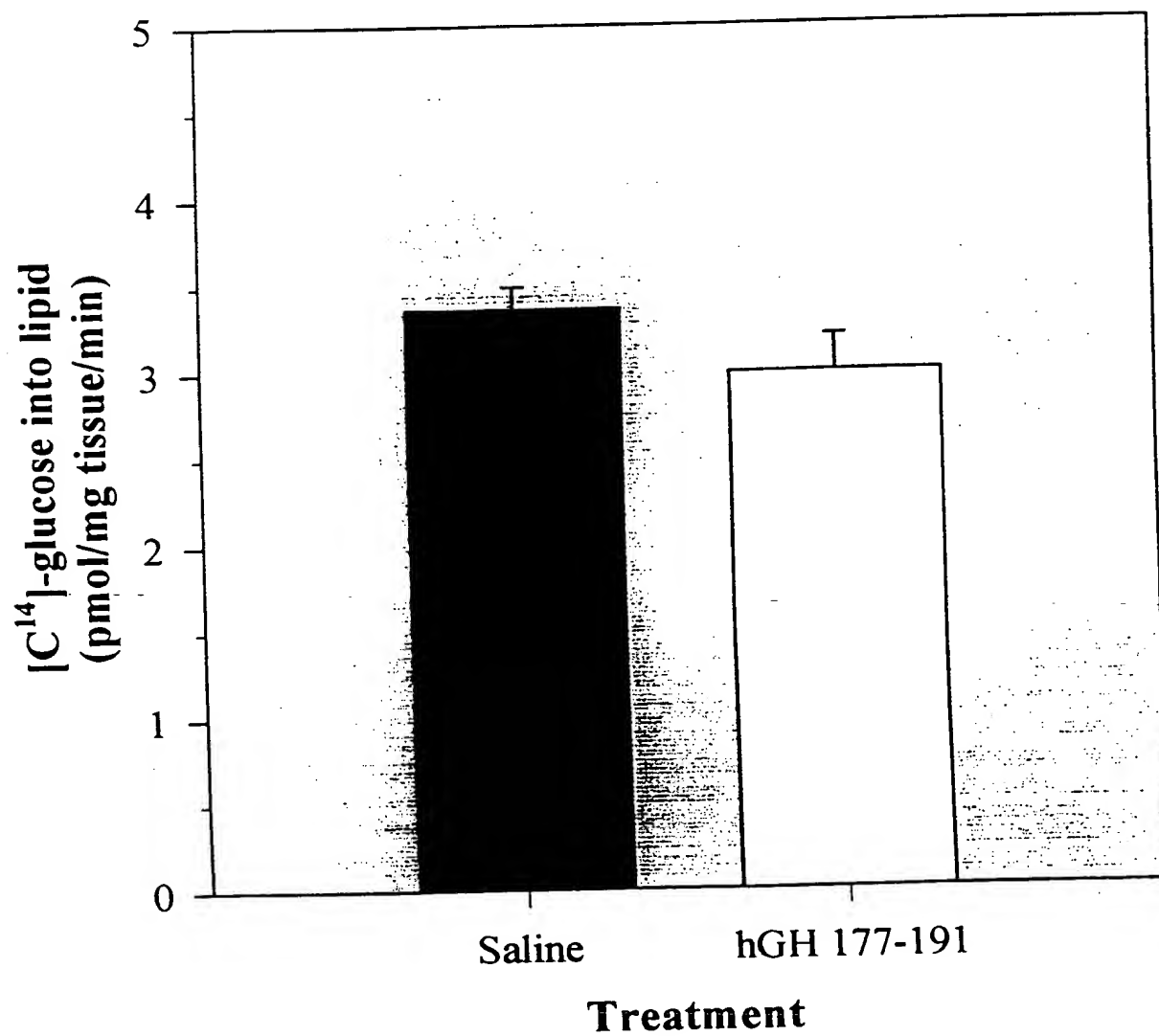


Figure 5B

***Ex vivo* effect on the lipolysis in adipose tissues of the male C57BL/6J (ob/ob) mice after 18-day Treatment**

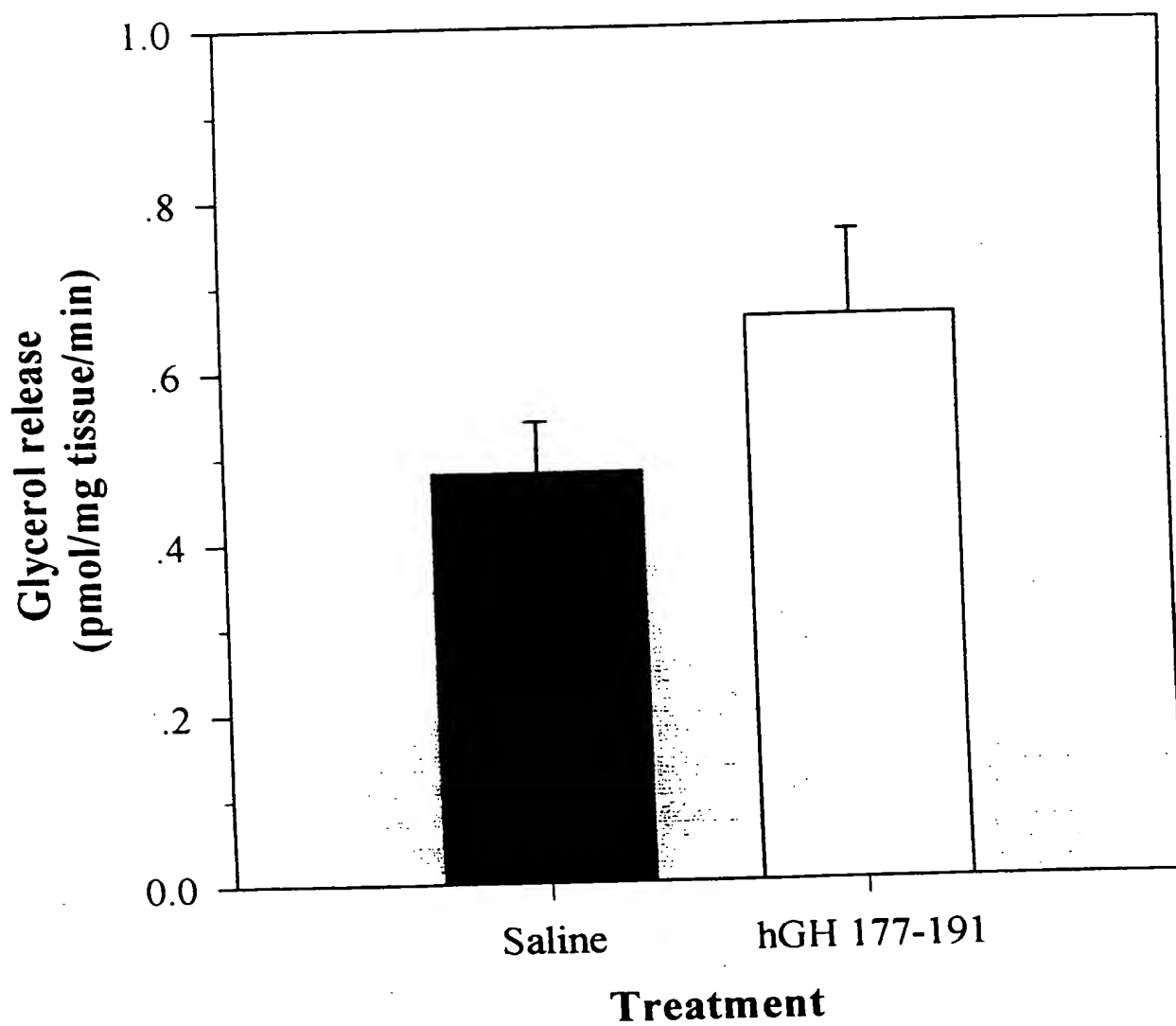


Figure 6A

***Ex vivo* effect on the lipolysis in adipose tissues of the female C57BL/6J (ob/ob) mice after 18-day treatment**

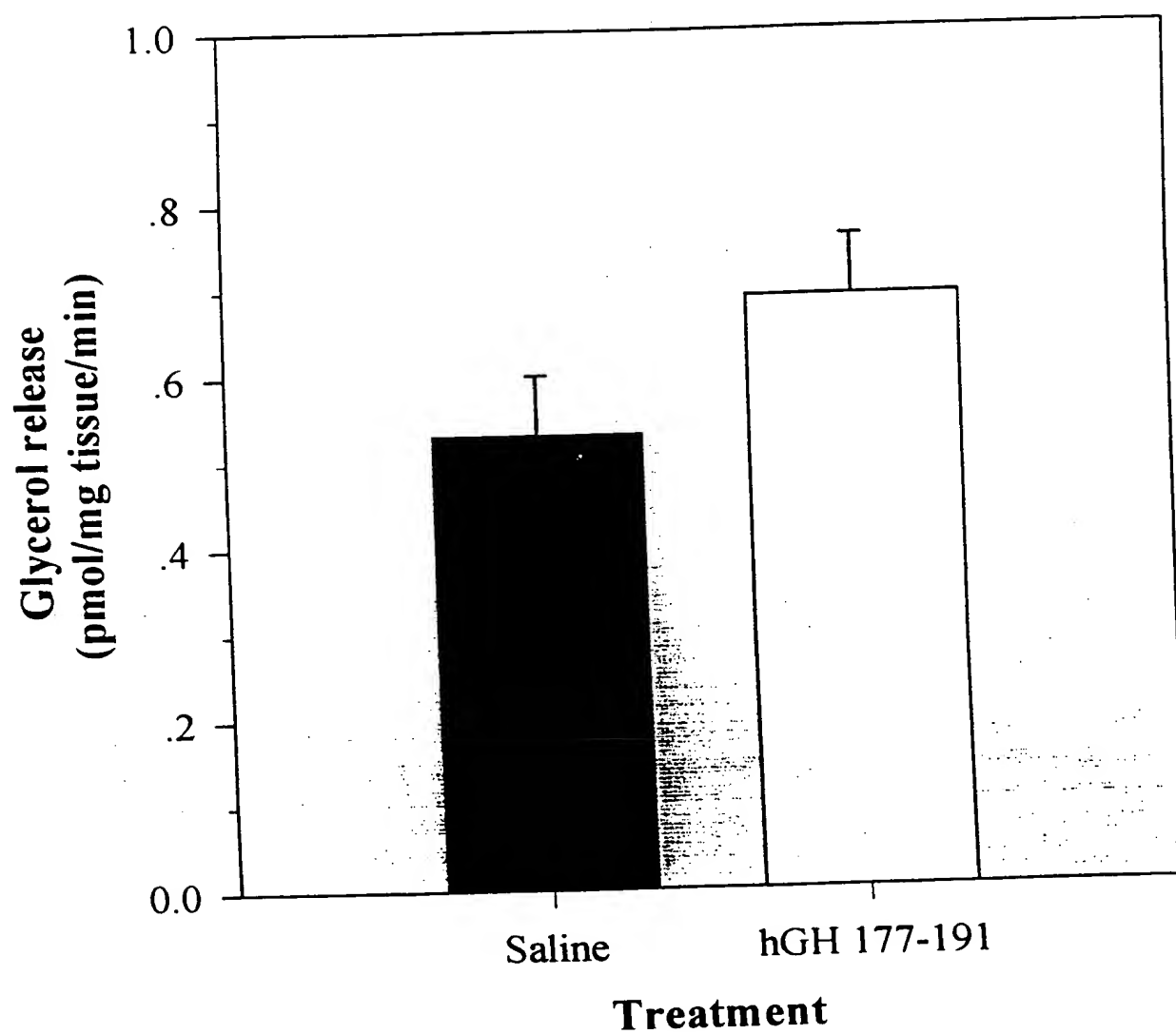


Figure 6B

In vitro effect of hGH 177-191 on fatty acid oxidation
in isolated adipose tissue of C57BL/6J (ob/ob) mice

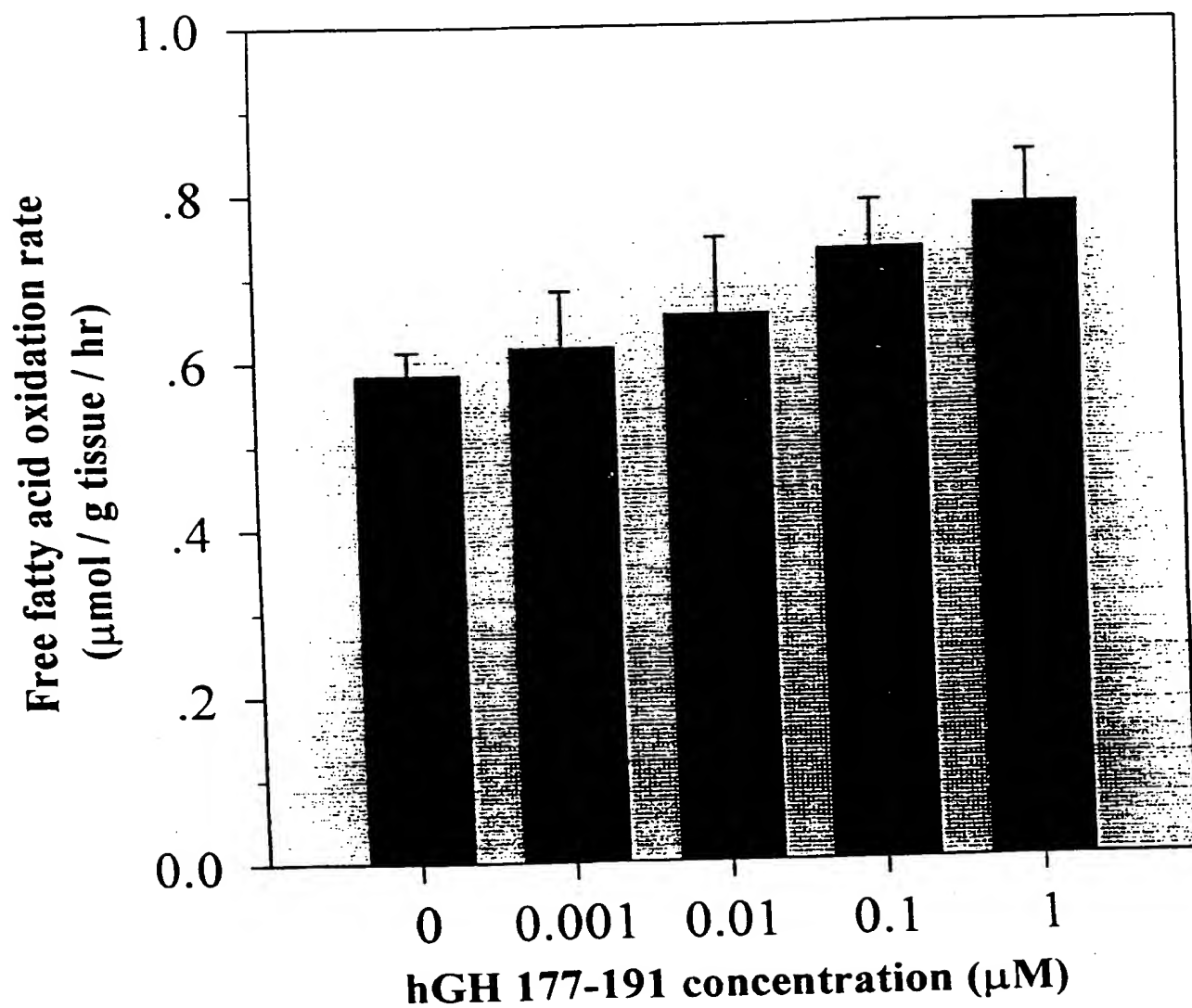


Figure 7

**Effect of hGH 177-191 on the diacylglycerol release
from isolated adipocytes**

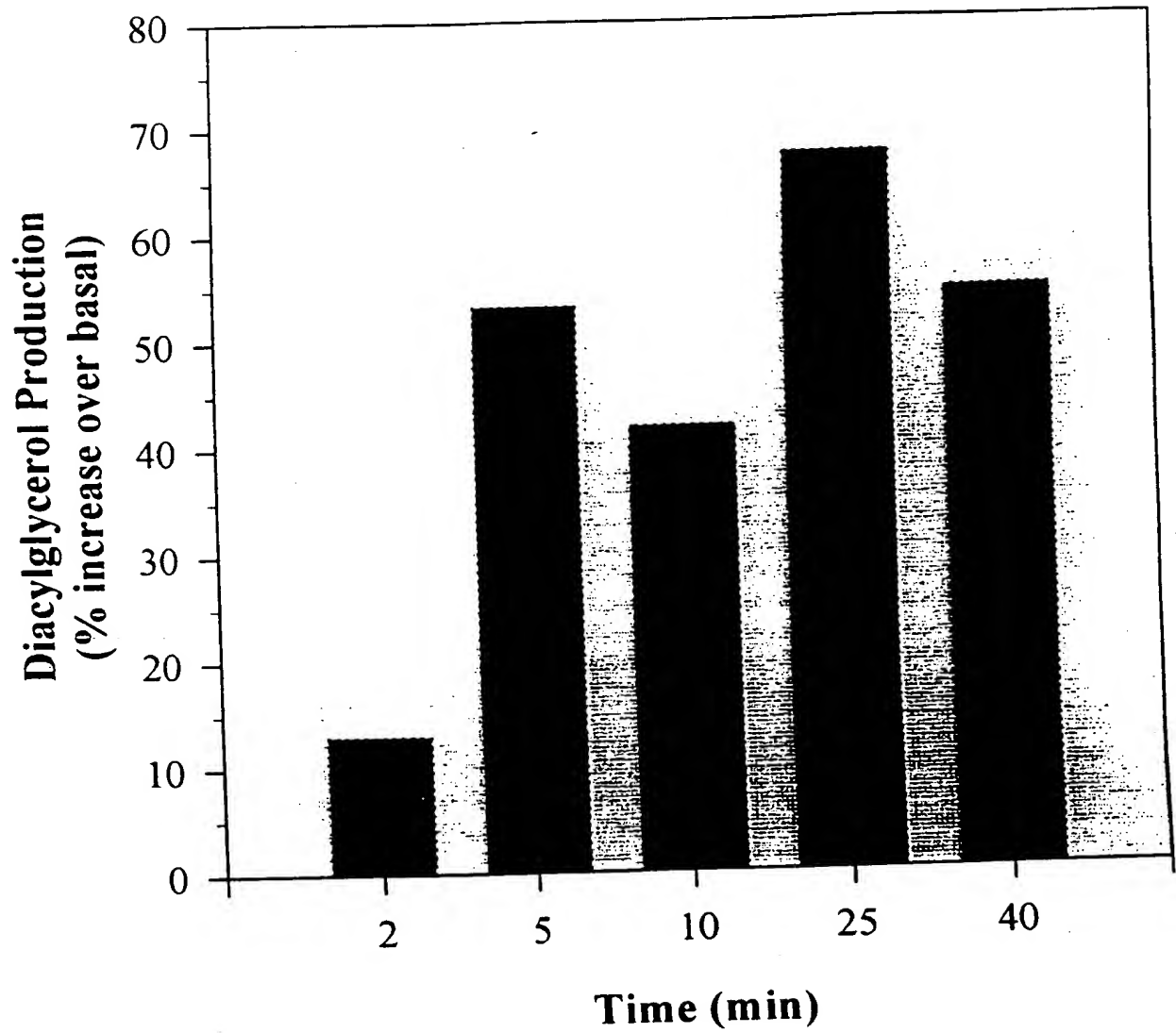


Figure 8

***In vitro* effect of hGH 177-191 on hormone-sensitive lipase activity in isolated adipocytes of male Zucker fatty (fa/fa) rats**

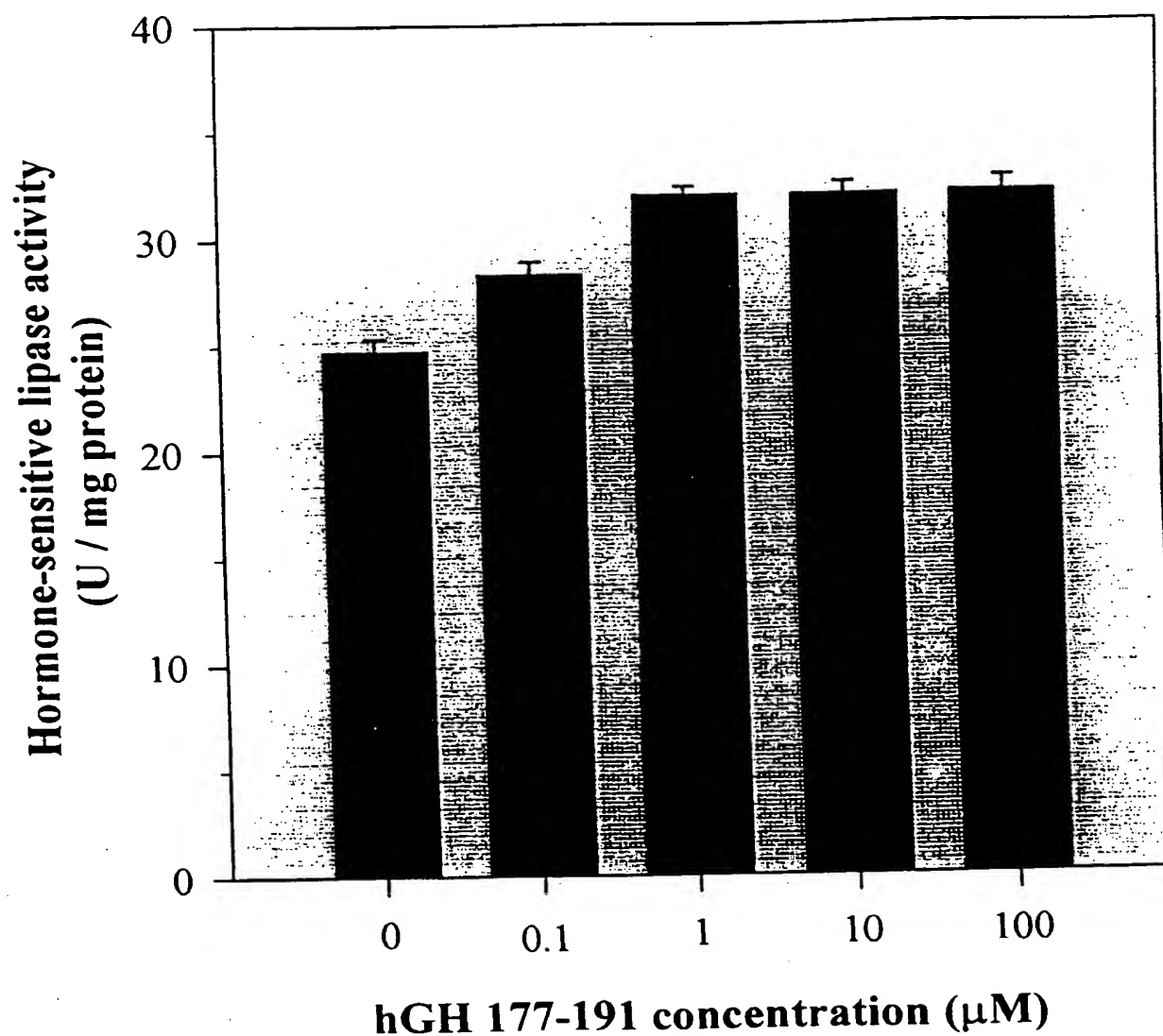


Figure 9

***In vitro* effect of hGH 177-191 on acetyl-CoA carboxylase
in the isolated adipocytes of male Wistar rats**

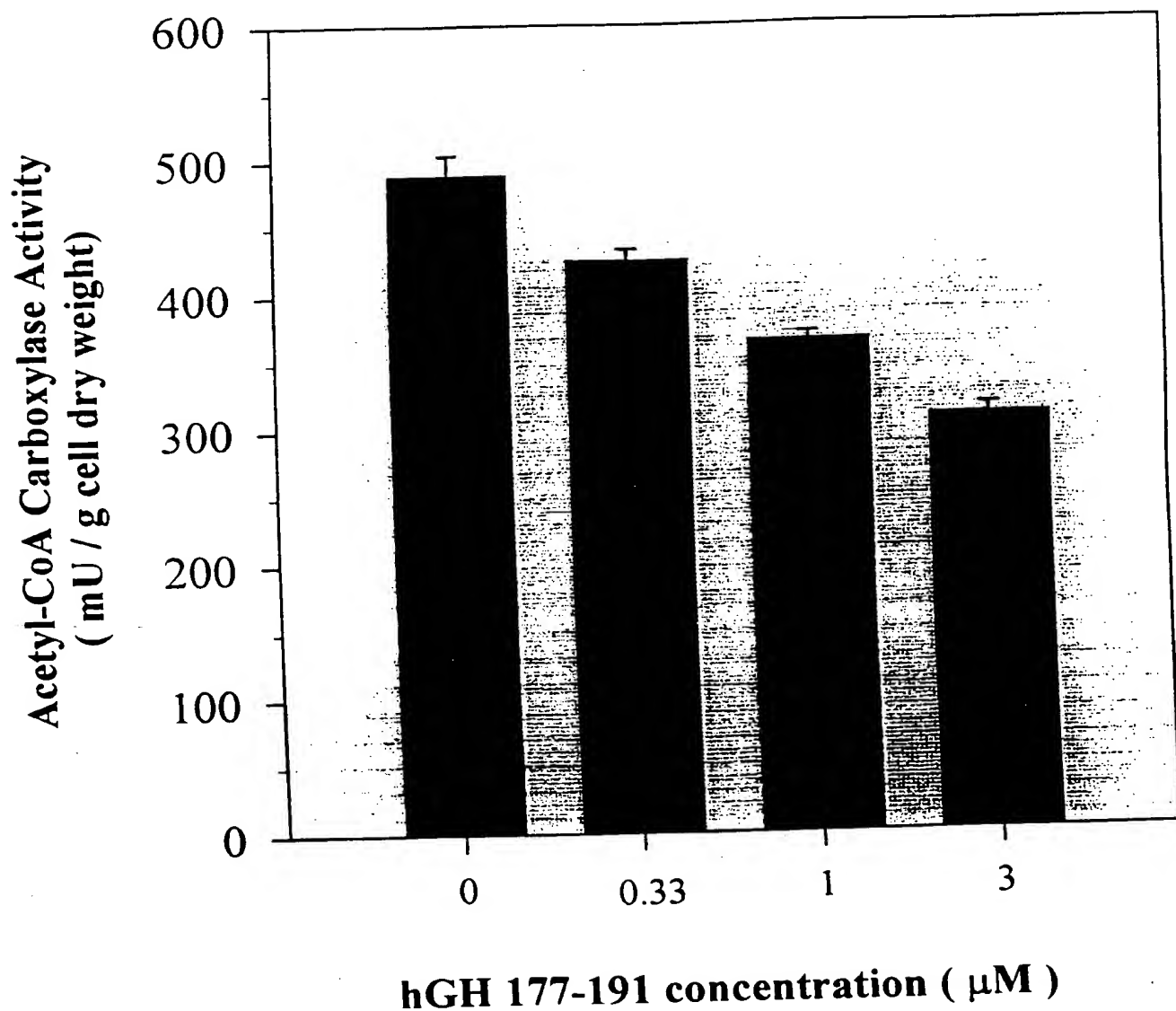


Figure 10A

In vitro effect of hGH 177-191 on acetyl-CoA carboxylase
in the isolated hepatocytes of male Wistar rats

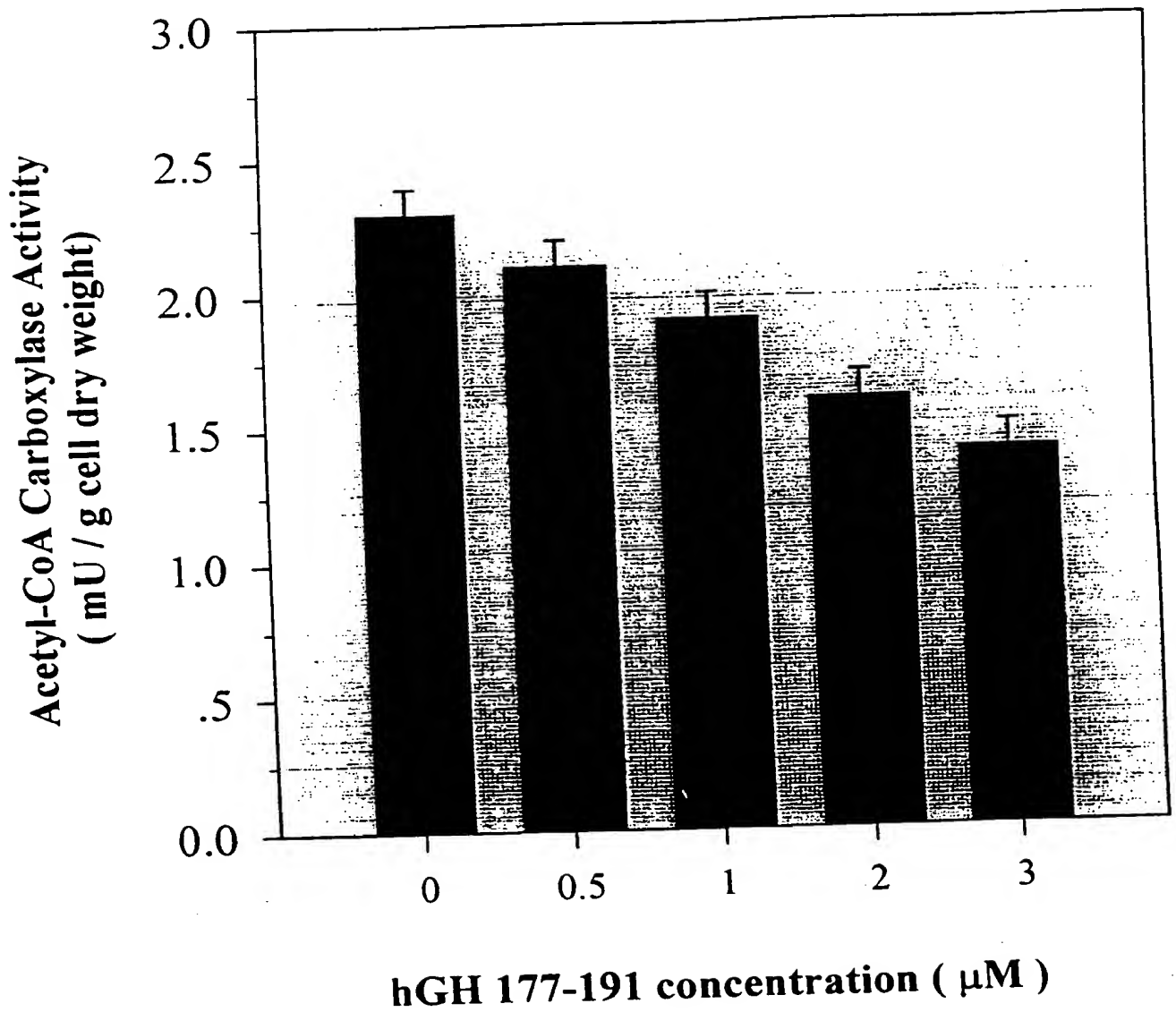


Figure 10B

Effect of chronic treatment on female C57 BL/6J (ob/ob) mice (26 week-old) by daily intraperitoneal injection with SEQ ID 11 (500 μ g/kg body weight)

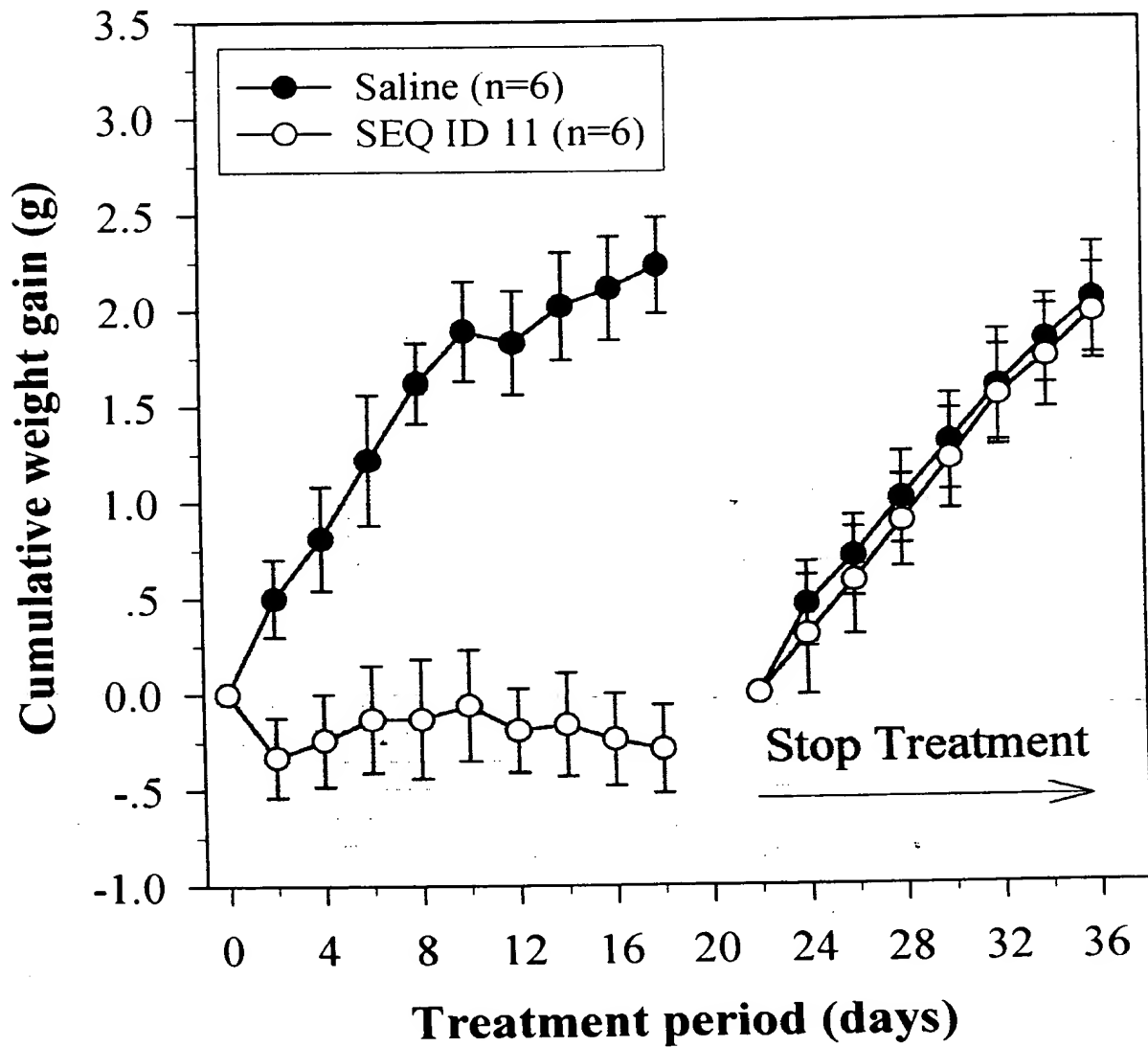


Figure 11A

Effect of chronic treatment on female C57 BL/6J (ob/ob) mice (26 week-old) by daily intraperitoneal injection with SEQ ID 11 (500 μ g/kg body weight)

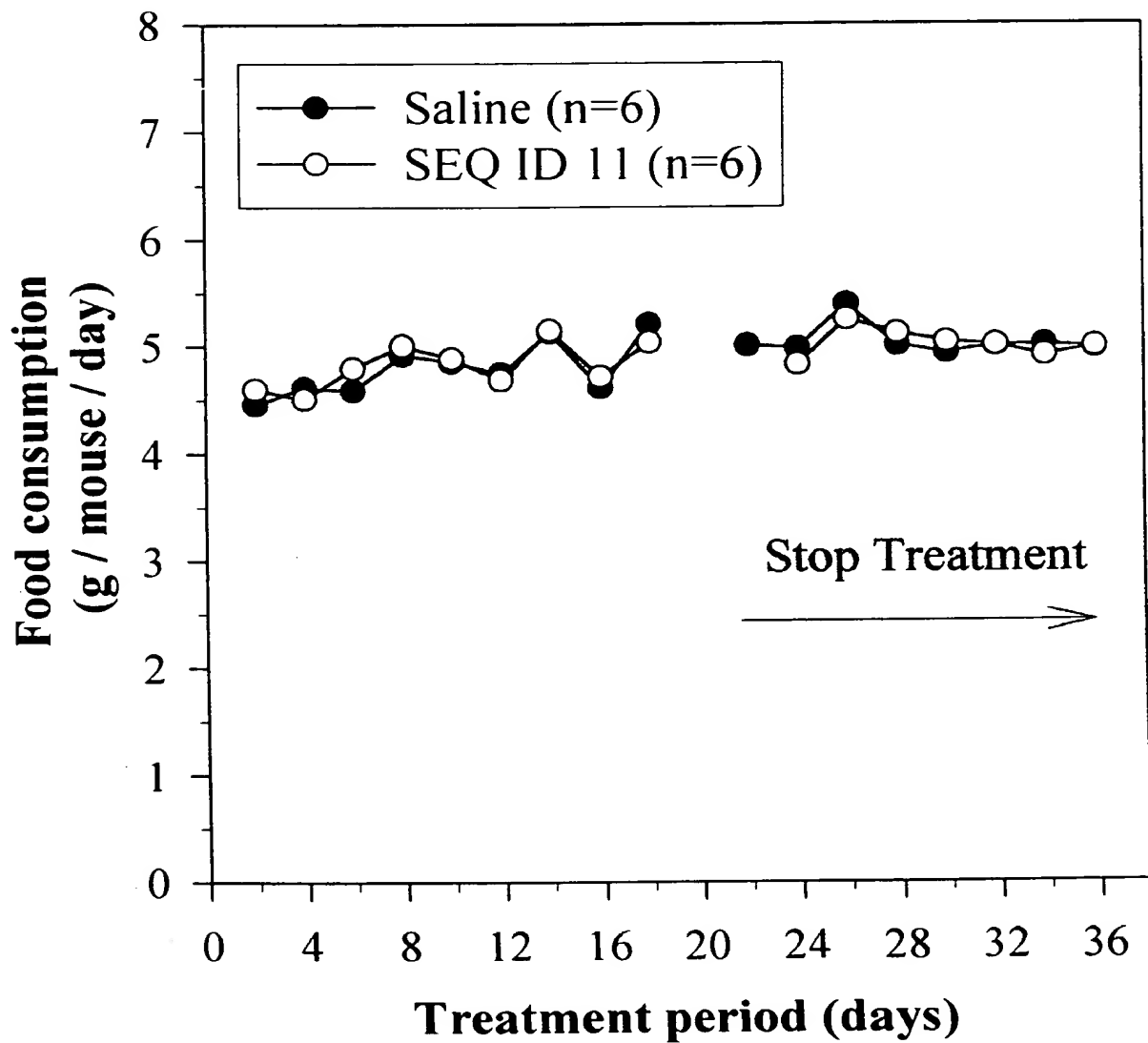


Figure 11B